TECHNICAL PUBLICATION 87-2

March 1987

DRE 233

PERIPHYTON AND WATER
QUALITY RELATIONSHIPS
IN THE EVERGLADES
WATER CONSERVATION AREAS
1978-1982

TECHNICAL PUBLICATION 87-2

March 1987

PERIPHYTON AND WATER QUALITY RELATIONSHIPS IN THE EVERGLADES WATER CONSERVATION AREAS 1978-1982

DRE 233

By David R. Swift Robert B. Nicholas

Environmental Sciences Division Resource Planning Department South Florida Water Management District West Palm Beach, FL

TABLE OF CONTENTS

<u>P</u>	age
List of Tables	i
List of Figures	i
Acknowledgements	
Executive Summary	
Introduction	1
Study Area	
Methods and Materials	_
Results	
Solar Insolation, Water Temperature	
Hydrology	
Major Ions	
Nitrogen and Phosphorus	
Periphyton Species Composition	
Geographical and Seasonal Distribution of Major Algal Groups.	
Periphyton Growth Rates	
Periphyton Nutrient Content	
Species Diversity	
Relationship between Periphyton Species Composition	23
and Environmental Factors	20
Discussion	
Major Ions, Algal Species Composition and Diversity	
Nutrients, Algal Growth Rates, and Nutritional Requirements	
Indicator Organisms	
Comparison of Peatland Marsh Water Chemistry	
Effects of Water Management on Everglades Periphyton	
Introduction of Nutrients as a Result of Backpumping.	
Increasing Marsh Water Ionic Content as a Result of Backpumping	
Literature Cited	41

Appendix A Species List Appendix B Taxonomic Notes

LIST OF TABLES

Table	<u>Page</u>	
1	Average major ion concentration and field measurements at interior WCA sites, February-June 1983.	11
2	Comparison of WCA-1 marsh surface waters with south Florida rainfall quality	13
3	A comparison of averaged nutrient concentrations at interior WCA marsh sites with Florida lakes and selected lake trophic state classification schemes.	14
4	Average nutrient concentrations at interior WCA sites, February 1977-June 1982.	15
5	Varimax rotated factor loadings for 30 biological and physiochemical variables	31
6	Correlation coefficients (R) from a stepwise multiple regression analyses of dependent biological variables vs. independent physiochemical variables.	32
7	Comparison of elemental N and P concentrations in algae and aquatic plants from different environments.	36
8	A comparison of average ionic composition of western European/Scandanavian peatland surface waters with averaged data from the three Water Conservation Areas	38
	LIST OF FIGURES	n
Figur	re	Page
1	Location of Everglades Water Conservation Areas andboundary of original Everglades in south Florida (adapted from Smith, 1968).	4
2	Location of long-term periphyton/water quality sampling sites in the Everglades Water Conservation Areas, 1978-1982	7
3	Light, water temperatures, and periphyton growth rates at the 217 gauge, WCA-2A, 1978-1982	10
4	Water levels, conductivity, and alkalinity, calcium and chloride concentrations at Site A-3, interior WCA-1 marsh, 1978-1982.	12
5	Water levels, conductivity, and alkalinity, calcium and chloride concentrations at the 217 gauge, interior WCA-2A marsh, 1978-1982.	13
6	Water levels, conductivity, and alkalinity, calcium and chloride concentrations at the 3-4 gauge, interior WCA-3A marsh, 1978-1982	14
7	Water levels, conductivity and alkalinity, calcium and chloride concentrations at the 3-7 gauge, interior WCA-3A marsh, 1978-1982.	15
8	Conductivity vs. hardness (Ca + Mg) in WCA surface waters, 1978-1982	17

LIST OF FIGURES (Cont'd)

Figur	re .	Page
9	Conductivity vs. chloride concentrations in WCA surface waters, 1978-1982	. 17
10	Seasonal variations in water level, nutrients, periphyton growth rates, and species composition at the 217 gauge, interior WCA-2A marsh, 1978-1982.	23
11	Seasonal variations in water level, nutrients, periphyton growth rates, and species composition at the 3-7 gauge, interior WCA-3A marsh, 1978-1982.	24
12	Seasonal variations in water level, nutrients, periphyton growth rates, and species composition at the 3-4 gauge, interior WCA-3A marsh, 1978-1982.	25
13	Seasonal variations in water level, nutrients, periphyton growth rates, and species composition at Site A-3, interior marsh, WCA-1, 1978-1982.	26
14	Seasonal variations in water level, nutrients, periphyton growth rates, and species composition at the 1-8T gauge, peripheral marsh, WCA-1, 1978-1982.	28
15	Seasonal variations in water level, nutrients, periphyton growth rates, and species composition at site B-2, peripheral WCA-3 marsh, 1978-1982	29
16	Seasonal variations in water level, nutrients, periphyton growth rates, and species composition at site B-5, peripheral WCA-2A marsh, 1978-1982.	30
17	Periphyton elemental N and P content at sites B-2, B-5 and the 217 gauge, WCA-2A	40
18a 18b	Periphyton elemental N and P content at sites A-3, and the 1-8T gauge, WCA-1	
19	Periphyton N:P mass ratio averages, standard deviations, and ranges at seven long-term WCA monitoring sites	43
20	Periphyton species diversity means, standard deviations, and ranges at seven long-term WCA monitoring sites.	45

ACKNOWLEDGEMENTS

This investigation was supported and conducted by the Environmental Sciences Division, Resource Planning Department, South Florida Water Management District, West Palm Beach, Florida.

The authors wish to express their appreciation to Dr. Benjamin F. McPherson, U. S. Geological Survey; Dr. Clinton Dawes, University of South Florida; Dr. Hugh Putnam, Water and Air Research, Inc.; and Dr. Patrick Gleason, James Montgomery Consulting Engineers, Inc., for their editorial review and comments.

EXECUTIVE SUMMARY

Submerged and floating mats of periphyton algae are a conspicuous feature of the Everglades Water Conservation Areas. These microscopic plant communities not only represent a major component of the marsh food chain, but also influence dissolved oxygen concentrations, calcium carbonate (marl) deposition, and nutrient cycling in the marsh. Although periphyton are recognized as an important component of the Everglades ecosystem, little information exists regarding their ecology, population dynamics, or relationship to chemical or physical parameters in the Water Conservation Areas. This is the second technical publication of a four year study (1978-1982) conducted by the SFWMD Environmental Sciences Division concerning algal research in the Water Conservation Areas. Results of the initial 1978-1979 study are available in Technical Publication 81-5 (Swift, 1981). Primary research questions studied in this project were: (a) how do environmental variables such as light, temperature, hydroperiod, or water quality factors affect periphyton species composition and growth in the marsh, (b) what effects do short term variations in water level or water quality have on periphyton community structure, and (c) how do present water management practices impact WCA periphyton populations? Results of this work will be useful in measuring rates of ecological change in the marsh over time and should serve as a baseline for assessing future water management alternatives within the study area.

A total of 333 species of algae representing 107 genera from 8 divisions (including 4 newly described species) were identified from glass slide substrates incubated in the three Water Conservation Areas. Water quality was the predominant factor which controlled periphyton species composition and growth. Hydroperiod and water level fluctuations played less important roles. Algal species composition and distribution at interior marsh sites were closely linked to ionic content and surface water origin. In contrast, peripheral marsh algal communities were strongly affected by nutrient enriched canal water inflows.

Surface water origin was the predominant factor regulating the mineral content of each marsh. Seasonal variations in mineral content were weakly correlated with rainfall and evapotranspiration. Surface waters within WCA-2A, WCA-3A, and the peripheral marsh of WCA-1 were highly mineralized

(calcium carbonate) and dominated by filamentous blue-greens (Schizothrix calcicola, Scytonema hofmannii) and an assemblage of hardwater diatoms (Mastogloia smithii v. lacustris, Cymbella ruttneri, Anomoeneis vitrea, and Synedra pahokeensis sp. nov.). Results indicate these algae are biological markers of calcium rich, low nutrient water quality conditions at interior marsh sites. In contrast, acidic, low ionic content waters (characteristic of interior WCA-1) supported a rich desmid-filamentous green (Mougeotia spp. and Spirogyra spp.) microflora.

Inorganic nitrogen and phosphorus concentrations at interior marsh sites were low in comparison to Florida lakes and peripheral marsh sites impacted by canal water inflows. Inorganic N and P remained low (near or below detection limits) when the marsh was flooded. During periods of marsh drawdown, inorganic nitrogen increased rapidly as waters receded. Phosphorus showed only minor increases in concentration during marsh drawdown and reflooding.

High concentrations of nitrogen and phosphorus were consistently recorded from two marsh sites (B-2 and B-5) located south of the S-10C structure in WCA-2A. These nutrient enriched sites supported a "specialized" periphyton community dominated by filamentous greens (Oedogonium spp. and Stigeoclonium sp.) in early summer, filamentous bluegreens (Microcoleus lynbyaceus) in late summer and fall, and diatoms during winter. The diatom flora present at these sites were pollution "indicator" species characteristic of nutrient enriched waters (eg. Gomphonema parvulum, Nitzschia amphibia, Navicula disputans, Navicula confervaceae, and Nitzschia palea).

Interior marsh periphyton communities showed a clear seasonal pattern of maximum growth from August through early October (end of wet season), and a period of diminution from January-March. Algal growth rates at interior marsh sites were low and were more affected by seasonal variations in light and temperature in comparison to seasonal changes in nutrient concentration. Highest growth rates recorded in the study occurred at nutrient enriched marsh sites (B-2 and B-5). Maximum and minimum growth rates at these sites were highly correlated with surface water phosphorus content.

Water quality and periphyton biomonitoring studies conducted in the northeast section of WCA-2A during 1980-82 showed nutrients to penetrate the marsh deeper than reported in the 1978-79 survey.

Supplies of elemental phosphorus in periphyton cell tissue from interior WCA-2A and WCA-3A sites were low in comparison to "critical concentrations" required for normal growth. Low supplies of phosphorus within marsh surface waters, periphyton cell tissue and low algal growth rates suggest nutrient limiting conditions at interior WCA-2A and WCA-3A sites. Elemental P concentrations at site A-3 (interior WCA-1) were 2-3 times higher than those recorded in WCA-2A and WCA-3A. Co-precipitation of phosphorus in calcium carbonate rich waters may partially explain the low concentration of elemental P observed in algal cell tissues collected from interior WCA-2A and WCA-3A marsh sites. Periphyton elemental P concentrations at peripheral marsh sites were significantly correlated with concentrations of phosphorus in marsh surface waters and high algal growth rates. Highest concentrations of elemental P in periphyton cell tissue occurred at sites B-2 and B-5, located south of the S-10C structure in WCA-2A.

Low ionic content waters (interior of WCA-1) were correlated with high algal species diversity and an abundant desmid flora. Periphyton species diversity in the interior of WCA-1 were significantly

higher than those recorded in WCA-2A, WCA-3A, and the peripheral marsh of WCA-1. Lowest diversity indices occurred in nutrient enriched waters (Site B-2).

Comparison of Everglades peatland water chemistry with European peat forming marshes classified Water Conservation Areas 2A and 3A as minerotrophic fens. Water quality in the interior of WCA-1 compared well with reported "transitional" stages of ecological succession from minerotrophic fen to the classic ombrotrophic "raised bog" community.

Changes in periphyton species composition as a result of pumping nutrient enriched canal waters across the marsh will increase algal biomass and reduce algal species diversity at the primary producer level. Changes in the base of the marsh food web may have far reaching impacts on the species composition and biomass of higher trophic level herbivores. Such effects might include: changes in food item size, nutritive value, or selection choices available to grazing invertebrates and fish. Simplification of the marsh food chain may ultimately reduce ecosystem stability. Discharging highly mineralized (calcium rich) canal water across marshes that historically contain low concentrations of dissolved minerals could significantly alter the algal flora and water quality characteristics of the marsh.

SUMMARY OF RELATIONSHIPS BETWEEN ENVIRONMENTAL VARIABLES AND PERIPHYTON COMMUNITY STRUCTURE

Water Quality Conditions

	Hard Water, High N & P	Hard Water, Low N & P	Soft Water, Low N & P
AREA	Peripheral Marsh, WCA-2A, and WCA-3A	Interior Marsh, WCA-2A, WCA- 3A, Peripheral Marsh WCA-1	Interior WCA-1
Dominant Periphyton Flora	Pollution tolerant species: e.g., Microcoleus lyngbyaceus, (blue-green); Oedoginium spp., Spirogyra spp. and Stigeoclonium spp. (greens); Gomphonema parvulum, Nitzschia amphibia, Navicula disputans, Nitzschia	Calcareous blue-green algae: Schizothrix calcicola, Scytonema hofmannii Hardwater diatoms: eg. Mastogloia smithii v. lacustris, Cymbella ruttneri, Anomoeneis vitrea	Desmids and filamentous greens; eg. Mougeotia spp. Oedogonium spp.; Diatoms indicative of acid, soft water; eg. Cymbella amphioxys, Frustulia rhomboides v. saxonica, Anomoeoneis serians v. brachysira
Seasonal Periodicity	Filamentous greens, spring/early summer; blue-greens, late summer and fall; diatoms, winter	Blue-greens; peak growth rates late summer and fall; diatoms winter/spring	Filamentous greens, summer; desmids, late summer, fall
Periphyton Growth Rates	High	Very low-low	Low
Algal Species Diversity	Low	Moderate	High
Cell Tissue Phosphorus Content	High (Luxury consumption of P)	Very Low (P Limited)	Low-moderate
Cell N:P Ratio	Low	High	Moderate-high
Effects of Light and Temperature	High water temperature and nutrients favor development of blue-greens (Microcolues sp.); low water temperature and nutrients favor filamentous green algae/diatom community	Light and water temperature important seasonal growth factors at interior WCA-2A and WCA-3A sites	Light and water temperature important seasonal growth factors at interior WCA-1 sites



INTRODUCTION

Submerged and floating mats of calcium carbonate encrusted algae, commonly referred to as periphyton, are a conspicuous and ecologically important element of south Florida wetlands. Periphyton are defined as an assemblage of attached microorganisms (primarily algae) which form living biofilms on the free surface of submerged substrates. As primary producers, periphyton potentially represent a major component of the marsh food chain. Periphyton mats provide both habitat and organic matter for grazing invertebrates and small forage fish which, in turn, are consumed by wading birds, sport fish, and reptiles (Tabb et. al., 1967; Carter et. al., 1973). Wood and Maynard (1974) estimate that a significant, if not greater, portion of the total primary production of the lower Everglades marsh is derived from the 200+ species of periphyton rather than from vascular plants. Similarly, Hunt (1961) estimated that nearly all the daily production and respiration of Everglades open water prairies is due to periphyton photosynthesis and respiration. Periphyton represent a primary food source for Everglades crayfish (Schomer and Drew, 1982 and references cited therein). Kolipinski and Higer (1969) report that filamentous blue-greens, desmids, and diatoms comprise a major portion of the diet of a number of Everglades forage fish.

In south Florida, periphyton mats seasonally cover large areas of the Everglades Water Conservation Areas. Periphyton photosynthesis and respiration markedly influence diel dissolved oxygen and carbon dioxide levels within the Everglades marsh (Hunt, 1961; Wilson, 1974). Periphyton growth and metabolism also influence nitrogen fixation (Goldstein et. al., 1980), marl deposition, soil building, and nutrient uptake processes within the marsh (Craighead, 1971; Gleason, 1972; Gleason and Spackman, 1974). Periphyton may also play an important role in the survival of aquatic organisms during drought. The formation of thick algal coatings on submerged substrates provides a moist microhabitat that prevents the dessication of eggs and larvae of insects, invertebrates, and fish during the dry season. As water levels rise, these organisms rapidly repopulate the reflooded marsh (Schomer and Drew, 1982 and references cited therein).

Until recently, studies of Everglades periphyton ecology have been largely restricted to Everglades National Park (Hunt, 1961; Brock, 1970; Van Meter-Kasonof, 1965, 1973; Gleason, 1972; Gleason and Spackman, 1974; Wood and Maynard, 1974; Wilson, 1974; Browder, 1981). Comparatively little data exists concerning the ecology of algal communities within the three Everglades Water Conservation Areas (Gleason and Spackman, 1974; Swift, 1981). Specific questions investigated in this study were: (a) Do

Water Conservation Area periphyton communities demonstrate periodic or seasonal changes in their community structure, growth rates, or nutritional requirements, (b) Are observed changes (if any) related to environmental variables such as light, temperature, hydroperiod, nutrients, pH, or major ion concentrations, (c) What effects do short term variations in water levels or water quality have on periphyton community structure, and (d) How do present water management practices impact Water Conservation Area periphyton?

This is the second phase of a program initiated in 1978 to study periphyton/water quality relationships within the three Water Conservation Areas. During 1978-1979, 18 periphyton/water quality sites were monitored. Water chemistry samples were collected concurrently with the growth of algae communities on glass slides over a sixty day incubation period (Swift, 1981). Results of these initial investigations suggested:

- (1) The distribution of major ions and nutrients varies greatly among sites and among Water Conservation Areas.
- (2) Nutrients are present in very low concentrations at interior marsh sites, suggesting nutrient limiting conditions.
- (3) Nutrient concentrations at marsh sites near canal water inflows were high in comparison to nutrient concentrations at interior marsh sites.
- (4) Periphyton species composition and growth rates on glass slides were significantly influenced by site differences in major ion content, pH, and phosphorus concentrations.
- (5) Calcareous blue-green algae and diatoms, characterized by low growth rates and low concentrations of elemental P, were the dominant algae at low nutrient, hard water interior marsh sites within WCA-2A, WCA-3A, and the peripheral marsh of WCA-1. Desmids and filamentous greens were the dominant algae within the acid, soft water interior marsh of WCA-1.
- (6) Periphyton communities exposed to nutrient enriched canal waters developed a "specialized" community of pollution-tolerant algae characterized by high growth rates and high concentrations of elemental P within algal cell tissues.
- (7) Phosphorus was shown to be the major water quality factor that controlled periphyton growth rates. Major ion concentration and pH appeared to heavily influence periphyton species composition at low nutrient, interior marsh sites.

Results of these preliminary efforts showed that further studies directed toward quantifying seasonal aspects of marsh water quality and periphyton ecology were warranted.

STUDY AREA

The three Everglades Water Conservation Areas (WCA) encompass almost a million acres (3500 km²) of native south Florida wetlands covering parts of Dade, Broward, and Palm Beach Counties. These fresh water marshes represent a little more than a third of the original 10,000 km2 Everglades ecosystem that once covered most of south Florida, extending from south of Lake Okeechobee 100 miles to the mangrove estuaries of Florida Bay and the Gulf of Mexico (Figure 1). Today, the Water Conservation Areas represent a modified wetland ecosystem with a network of canals, levees, pumps and gates which enclose the marsh allowing manipulation of water levels for flood control, water storage, groundwater recharge, wildlife management and recreational benefits. Water levels within the WCA are managed by the South Florida Water Management District.

The Everglades region has been described as a vast sawgrass (Cladium jamaicense) marsh dotted with tree islands and interspersed with wet prairies and aquatic sloughs (Davis, 1943, 1946). Poor drainage and decomposition of marsh vegetation in this lowland region have produced muck and peat deposits up to 4 m in depth that overlie calcareous limestone formations (Stephens, 1956; Gleason et. al., 1974). South Florida has a subtropical climate with generally frost free winters and long, hot, humid summers. More than 75 percent of the annual rainfall occurs during the June through October wet season. Water levels within the Water Conservation Areas fluctuate on a seasonal basis in response to rainfall and water management practices. Maximum water depths within the marsh typically range from 0.5-0.9 m during the late wet season (September-October) while some areas of the marsh are dry during March-April.

The majority of water that flows into the WCA's is derived from agricultural lands (sugarcane, rice, vegetable and sod farming, improved pasture). Canal waters, which drain agricultural lands, are rich in organic and inorganic forms of nitrogen and phosphorus and are highly mineralized due to mixing with ground water inflows (Gleason, 1974; Waller and Earle, 1975; McPherson et. al., 1976; Lutz, 1977; Millar, 1980).

Water Conservation Area 1, (WCA-1) also known as the Loxahatchee National Wildlife Refuge, is the northernmost pool of this modified wetlands system and occupies an area of 572 km² in western Palm Beach County. Wildlife and recreational resources of WCA-1 are managed by the U.S. Fish and Wildlife Service. WCA-1 is a relatively shallow marsh encircled by a rim canal that is 3 to 4 meters deep. Nutrient enriched canal waters derived predominantly from the Everglades Agricultural Areas and

grazing lands to the north are pumped into WCA-1 via S-5A and S-6 (Figures 1 and 2). During low water periods, these waters generally flow south along perimeter canals and do not penetrate the interior marsh due to slightly higher ground level elevations at the center of WCA-1. Under these conditions water quality within the marsh interior reflects that of rainfall. In the wet season, marsh stage levels rise in response to rainfall and increased pumping activities. High stage levels allow canal waters to penetrate the peripheral marsh to a much greater extent (Millar, 1980).

Water Conservation Area 2A (WCA-2A), the smallest of the three areas, lies to the southwest of WCA-1 and covers an area of 547 km² in southern Palm Beach and northern Broward Counties. WCA-2A receives large quantities of nutrient enriched canal water derived from the Everglades Agricultural Areas through the three S-10 discharge structures located on the Hillsborough Canal (Figure 2). The absence of interior canals along the north and east levees of WCA-2A forces this nutrient enriched water to sheet flow across the marsh. construction of canals in 1961, nutrient enriched S-10 water has impacted approximately 2,400 ha of marsh in the northeast portion of WCA-2A, producing significant changes in the species composition of aquatic macrophytes and periphyton (Gleason, 1974; Davis and Harris, 1978; Swift, 1981).

Water Conservation Area 3A (WCA-3A), located in western Broward and northwestern Dade Counties, is four times (2,400 km²) larger than WCA-2A. WCA-3A is bisected by interior canals so that the north end receives surface water runoff from agricultural lands (L-3, S-8, S-150). The S-9 structure pumps predominantly urban runoff into WCA-3A from the South New River Canal. The L-28E and S-140 drainage basins derive water from grazing lands and wetland communities located to the west of WCA-3A.

METHODS AND MATERIALS

In the first phase of this study, water chemistry samples were collected during the wet and dry seasons of 1978-79 concurrently with the growth of periphyton communities on glass slides at 18 sites located throughout the WCA system (Swift, 1981). During 1980-82, seven of these original 18 sites were sampled on a monthly basis (Figure 2).

Field data (temperature, dissolved oxygen, specific conductance, and pH) were recorded using a Hydrolab^R electronic recording instrument. Water quality samples were filtered in the field (Millipore^R membrane filter, pore size 0.45 microns) and stored in polyethylene bottles on ice. Major ions (sodium, potassim, calcium, magnesium, silicate, and total

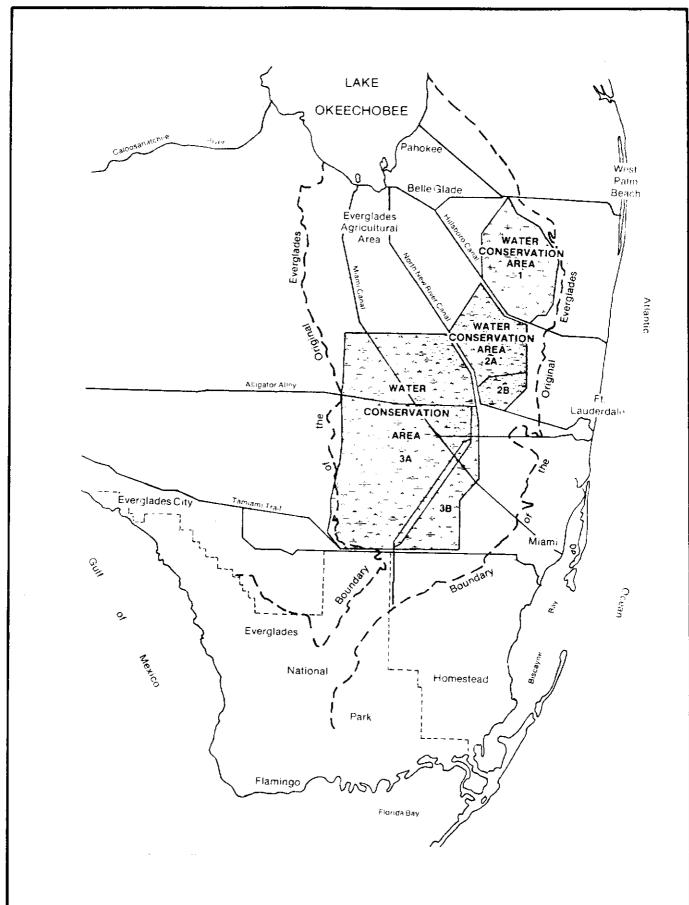
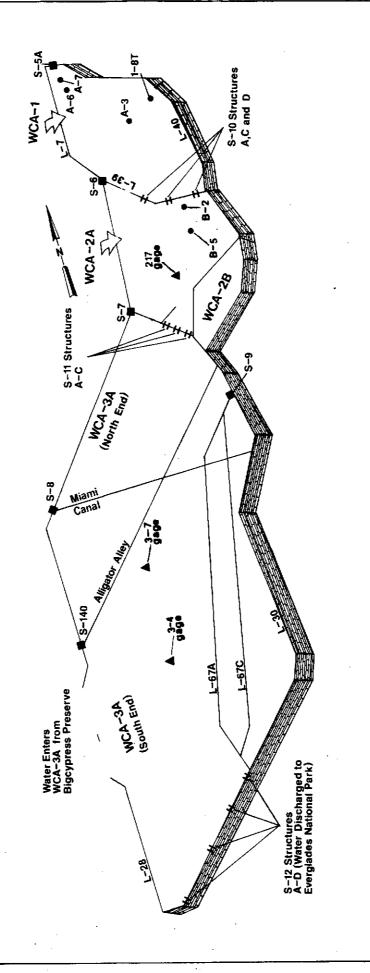


FIGURE 1. LOCATION OF EVERGLADES WATER CONSERVATION AREAS AND BOUNDARY OF ORIGINAL EVERGLADES IN SOUTH FLORIDA (Adapted from Smith, 1968).



LOCATION OF LONG-TERM PERIPHYTON/WATER QUALITY SAMPLING SITES IN THE EVERGLADES WATER CONSERVATION AREAS, 1978-1982. FIGURE 2.

● -Periphyton / Water Quality Sampling Sites ■-Pump Stations エ- Water Control Structures iron) were determined using flame atomic emission, while major nutrients (Total N, NH₄+, NO₃, NO₂, Total PO₄, total dissolved PO₄, and ortho-PO₄), chloride, sulfate, and alkalinity were measured using Technicon^R AutoAnalyzer methods. These chemical methods were recommended or approved by the Environmental Protection Agency (EPA, 1976) or the American Public Health Association (APHA, 1975).

Rainfall and surface water elevations were obtained from U.S. Army Corps of Engineers gauging stations located near the field sampling sites. Water depths were measured at the time of sample collection and correlated with nearby gauge data. Soil (peat) depths were measured at each site with a steel rod. Solar radiation data were obtained from the Agricultural Research and Education Center located near Belle Glade, Palm Beach County, Fla. (Lat. 26°40', Long. 80°38'). Data were reported in langleys (gram calories per cm²) measured by a solar pyranometer (Allen, 1978-82).

Periphyton communities were grown on glass slides suspended in redwood slide racks 3 cm below the water surface. Slides were collected from the wood racks every 30-35 days. Samples were analyzed for chlorophyll a content, species composition, relative abundance of major algal groups, cell density, cell volume, and species diversity based on the Shannon-Weaver equation (Shannon-Weaver, 1963). Periphyton communities growing on natural substrates were collected from each site and analyzed for elemental concentrations of N and P according to methods outlined by Black (1965). Each of the above methods are described in detail in an earlier report (Swift, 1981). Algal species identifications were performed by the authors. New species were confirmed by Dr. Charles Reimer, Academy of Sciences of Philadelphia.

Factor analysis was used as a screening technique to uncover underlying factors which account for the majority of variance contained within the data matrix. The method of factoring used was a principal component solution with Varimax-rotation with a minimum acceptable eigenvalue of 1.0. Tests of relationships between variables were also conducted using a stepwise multiple regression statistical package (Systems/360 IBM Subroutine Package). Percent relative abundance data based on cell volume estimates were arcsin transformed (Snedecor and Cochran, 1967) for use in the factor and multiple regression analyses.

RESULTS

Solar Insolation and Water Temperature

Maximum water temperatures reached 35.5°C on July 7, 1978 with the lowest temperature of 13.2°C

recorded on February 28, 1978 and January 10, 1979. The highest single daily solar radiation value was 716 langleys recorded on May 11, 1982, and the minimum value, 37 langleys, occurred on December 28, 1978 (Allen, 1982). Water temperature and averaged biweekly solar radiation showed typical, seasonal bell-shaped curves with maximum values occurring in June-August and minimum values occurring from December-February (Figure 3). Seasonal maximum water temperatures appeared to lag behind maximum light intensities by 1-3 weeks.

Hydrology

Marsh hydrology played an important role in determining seasonal variations in mineral and nutrient content of marsh surface waters at interior marsh sites (Figures 4-7).

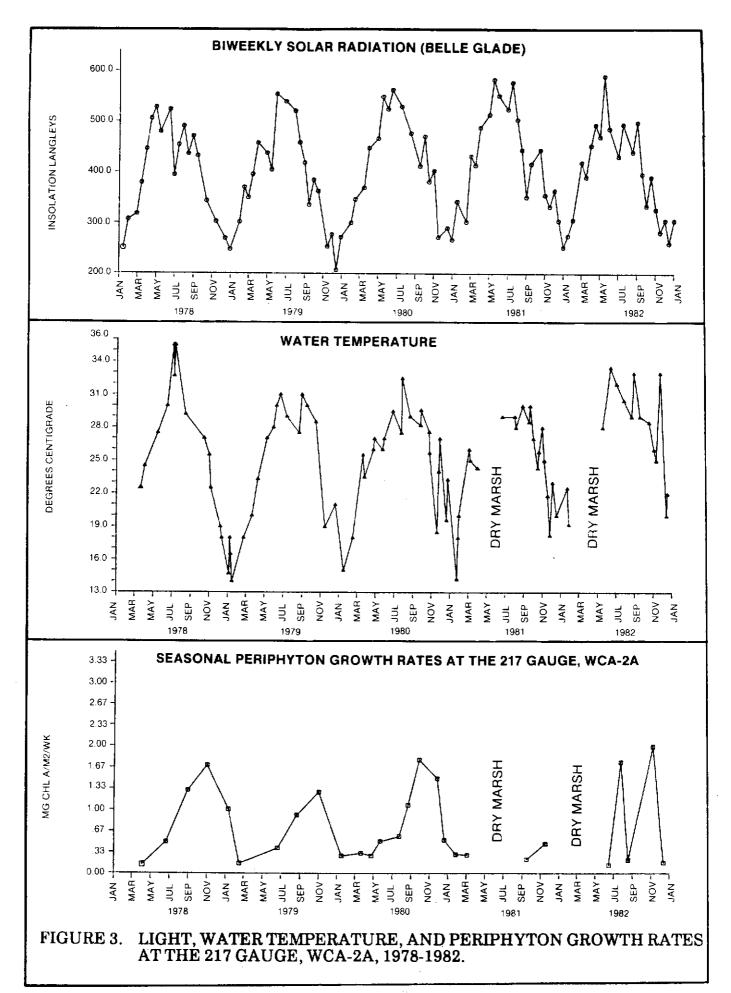
In WCA-1, station A-3 experienced the most prolonged hydroperiod. Water levels remained above ground level except during one brief dry period in April 1981 (Figure 4). Highest water levels occurred in September 1982 with a depth of 70 cm. Water levels generally decreased during winter and spring months (dry season) with rising stages occurring from mid-summer through fall (wet season).

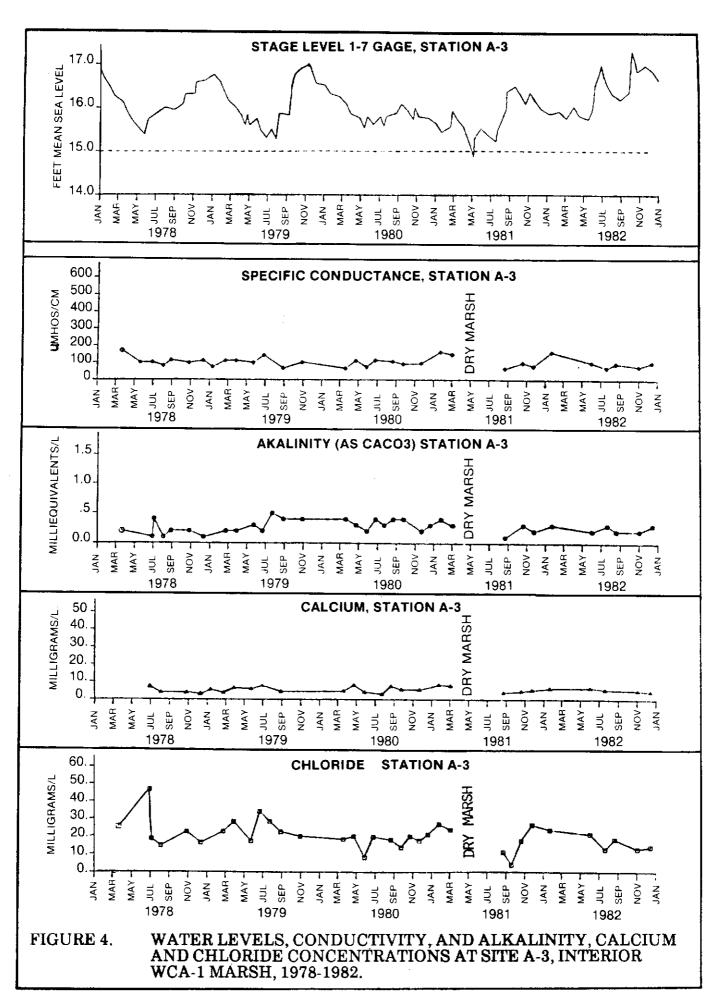
Water levels in WCA-2A remained well above ground from 1978 until April 1981. Figure 5 reflects the storage of water in WCA-2A (217 gauge) during 1978-79 with minimum and maximum depths of 0.36 and 1.10m, respectively. The experimental drawdown of WCA-2A during the 1981 drought caused water levels to fall well below ground level during April-May of that year. A less severe drying of the marsh occurred during January-April of 1982 (Worth, 1983).

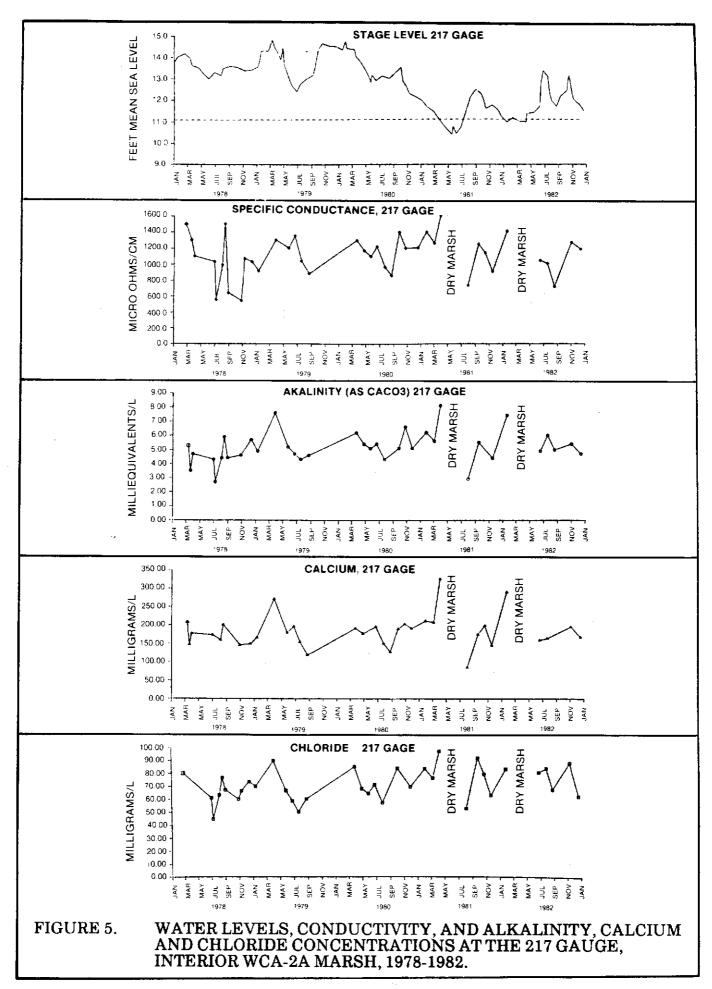
Water level fluctuations in WCA-3A were more typical of south Florida wetlands with distinct seasonal wet and dry cycles occurring each year (Figures 6 and 7). During 1981 and 1982, water levels in the northern portion of WCA-3A (3-7 gauge) fell to 0.8 m below ground level for periods of 5-6 months. Due to the low water table, numerous fires swept the area. Many of these burns were peat fires that caused extensive soil subsidence and loss of tree island communities.

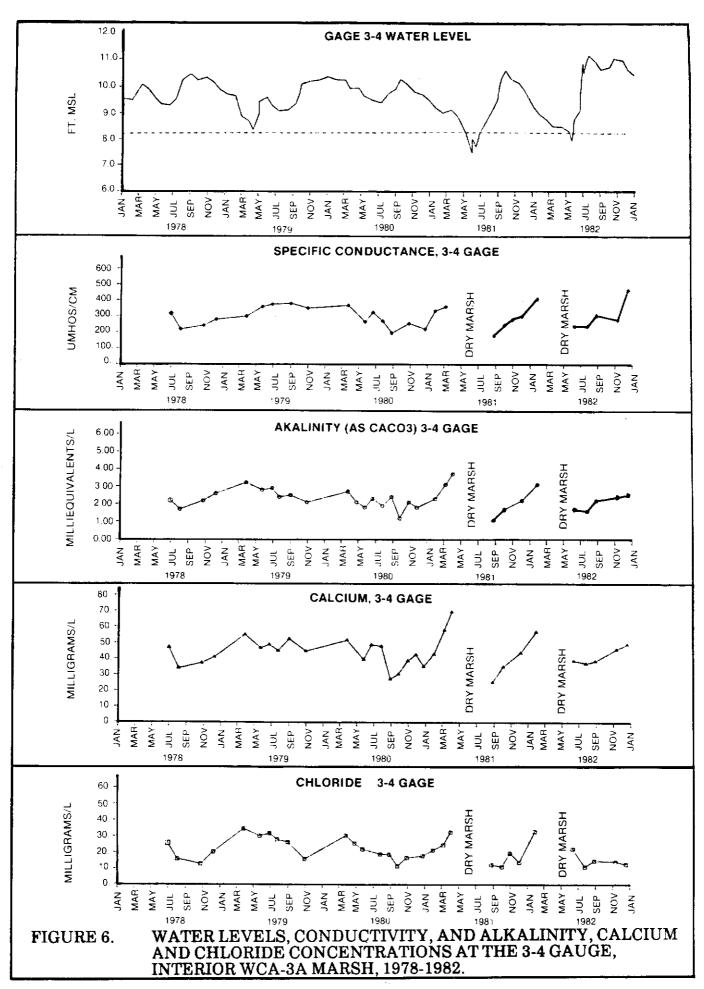
Major Ions

Interior marsh surface waters differed greatly in specific conductivity and mineral content among the three WCA's (Table 1). Lowest values of conductivity and major ion concentration occurred in the acid, soft water interior marsh of WCA-1, while highest conductivity and major ion values occurred in the highly mineralized waters of WCA-2A (Figures 8 and 9).









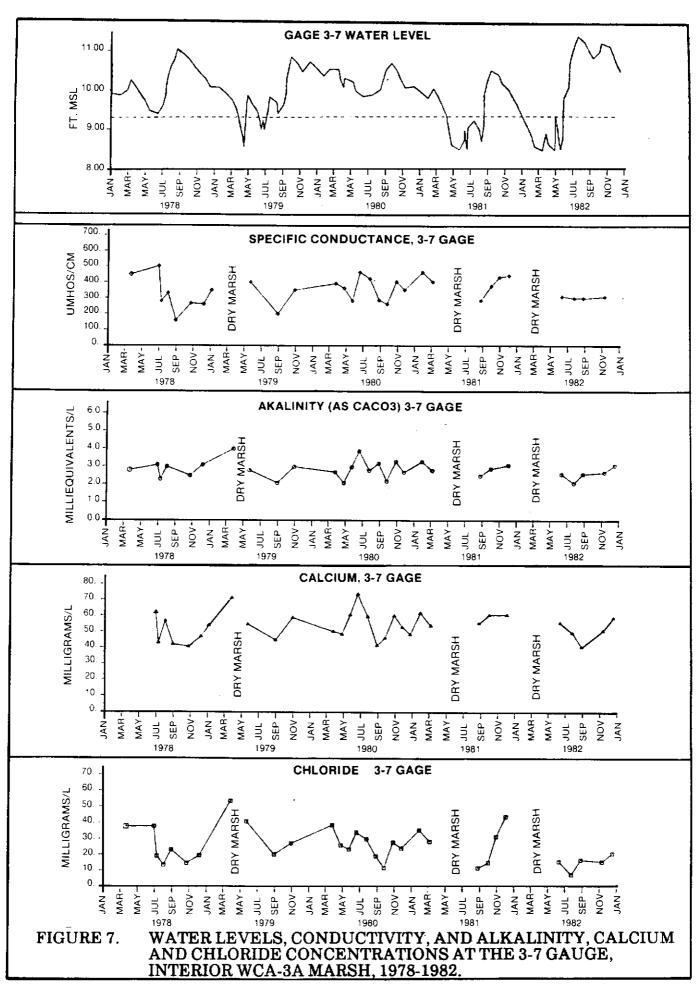


TABLE 1.	AVERAGE MAJOR MENTS AT INTERIO			
Parameter Field Measu	All Si Combi <u>(Mea</u> rements:	ned WCA-		-
. ICIG IVICASA	- Circuitor			
pΗ	6.8 (.7)		7.0 (.5)	7.1 (.4)
Conductivity µmhos c ^{m-1}	y 535 (403		333 (115)	1050 (269)
D.O. (mg/l)	5.3 (2.7		6.1 (2.2)	3.7 (2.3)
Major lons				
HCO₃ ⁼	177 (115		161 (34)	299 (61)
Alkalinity (as CaCO ₃)	145 (94		132 (28)	245 (50)
Hardness	155 (103		137 (30)	284 (57)
CL.	76 (72		28 (19)	174 (49)
Na⁺	49 (54		18 (10)	121 (30)
Ca⁺	45 (2 6		47 (9)	67 (14)
Mg⁺⁺	11 (11		4 (2)	28 (6)
K*	2.7 (2.6		1.1 (.9)	6.5 (2.1)
SiO ₂	10 (7)		7 (4)	19 (5)
SO ₄ =	18 (25		12 (27)	39 (19)
TOTAL-Fe	.3 6.)		.5 (.6)	.1 (.1)
CI:Ca Ratio	1.7	7 3.7	0.6	2.6
Upper value	e = avg; lower value	in parentheses	= standard devi	ation

The high ionic content of these waters are the result of highly mineralized agricultural water discharged into WCA-2A via the three S-10 structures located on the Hillsborough canal (Figure 1).

In contrast, surface waters in the interior of WCA-1 (Site A-3) were low in dissolved minerals, poorly buffered, acid and soft (Table 1). Specific conductivity at Site A-3 averaged 112 umhos/cm with pH in the acid range (4.6-6.8). Chloride (avg. = 22 mg/l), bicarbonate (avg = 19 mg/l), and sodium (avg. = 12 mg/l) were the dominant ions. Major ion concentrations in these waters were only 2-3 times higher than south Florida rainfall (Table 2) and suggest that rainfall is the primary source of water for the interior WCA-1 marsh.

Alkalinity and hardness of surface waters in the interior of WCA-3A were roughly one half the mineral content of WCA-2A. Specific conductance ranged between 300-450 µmhos/cm. Bicar-bonate (avg. = 161 mg/l) and calcium (avg. = 47 mg/l) were the dominant ions.

Seasonal variations in marsh water mineral content were more closely related to the origin of each marsh's water supply in comparison to factors such as rainfall.

Water quality within the interior of WCA-2A was highly mineralized with bicarbonate (avg. = 299 mg/l) and chloride (avg. = 174 mg/l) being the principal cations. Sodium, calcium, sulfate, and magnesium were the primary anions. Specific conductance ranged from $800\text{-}1300 \text{ }\mu\text{mhos/cm}$ and pH was within the circumneutral to basic range (6.8-7.5).

Linear regression results showed only weak correlations between water levels and marsh water conductivity or hardness over time. However, highest major ion concentrations were usually recorded during low water periods, while minimum concentrations occurred near the peak of the wet season (Figures 4-7).

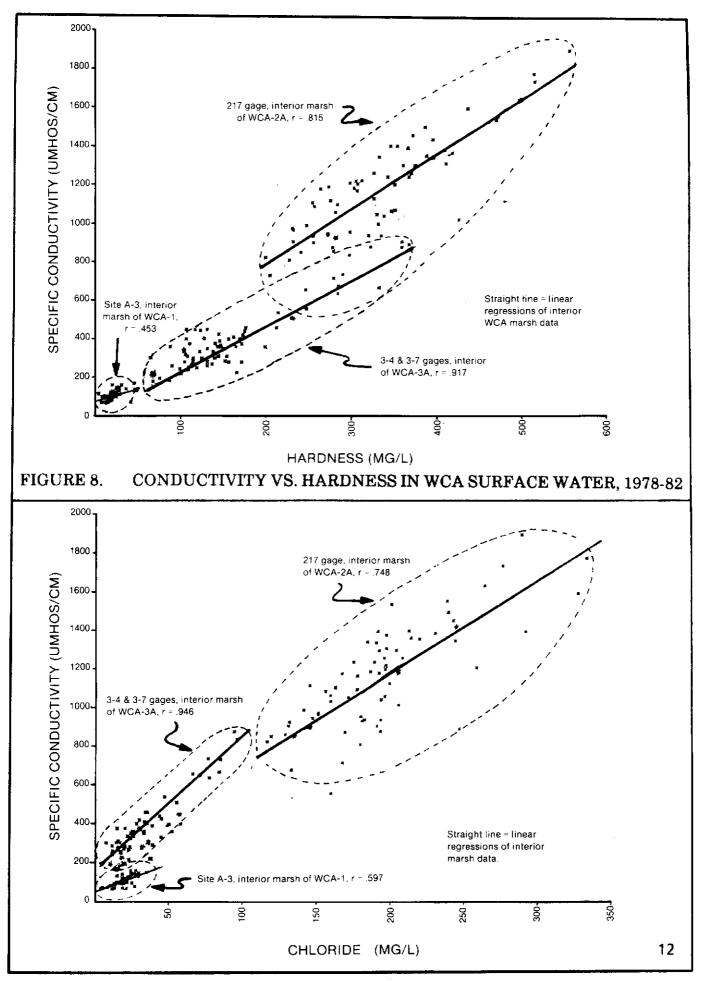


FIGURE 9. CONDUCTIVITY VS. CHLORIDE IN WCA SURFACE WATERS, 1978-82.

TABLE 2. COMPA	RISON OF	WCA-1	MARSH	1 SURF	ACE WA	ATERS V	VITH SO	UTH FLO	ORIDA F	RAINFAI	LL
Rainfall Collection Site	pH (Range)	Spec. Cond	<u>Ca</u> + +	<u>Cl</u> -	<u>Na</u> +	<u>K</u> +	<u>Mg</u> ++	<u>HCO</u> 3*	CaCO ₃ Hard- ness	Total <u>N</u>	Total P
Florida State- wide*Average	4.7 - 8.8	32	2.1 (0.8)	3.2 (2.0)	1.7 (1.3)	-	-	-	-	1.1 (.52)	.10 (.08)
S-5A at Loxa- hatchee*Florida	5.6- 6.4	41 (31)	3.4 (2.0)	3.9 (2.9)	2.3 (2.2)	.4 (.3)	.6 (.5)	5.6 (4.2)	11 (6)	.97 (.45)	.09 (.13)
S-9, Andytown* Florida	4.7- 6.6	66 (37)	2.9 (1.2)	7.9 (5.3)	4.4 (2.8)	.6 (.6)	.6 (.2)	4 (4)	9 (4)	2.4 (1.5)	.10 (.08)
Surface Water WC	CA-1 Interi	or Mars	<u>h</u>								
Average of ThreeSites (A-3, A-2, 1-9)	4.6- 7.1	112 (39)	6.3 (2.2)	22.1 (8.3)	12.3 (4.2)	.6 (.4)	1.5 (.7)	19.5 (9.1)	21 (9)	2.79 (3.95)	.015 (.029)

^{*}Source: Irwin and Kirkland, 1980, U.S. Geological Survey

Upper value = mean; value in parentheses = standard deviation

All values mg/I with the exception of pH and specific conductivity (µmhos/cm)

Seasonal variations in major ion content at peripheral WCA-2A marsh sites were significantly influenced by canal water inflows from the S-10 structures. Sites B-2 and B-5 showed significant correlations between rising water stages, chloride content, and nutrient concentrations (R=.690) as a result of their close proximity to the S-10 structures.

Nitrogen and Phosphorus

Average total PO₄, total dissolved PO₄, ortho-PO₄ and inorganic nitrogen concentrations at interior marsh sites were low in comparison to Florida lakes (within the oligotrophic range) and various lake trophic state classification schemes (Tables 3 and 4). Seasonal changes in inorganic nitrogen concentrations at interior marsh sites were closely related to marsh hydrology and, as a result, varied from year to year (Figures 10-13). Concentrations of inorganic nitrogen were at, or very near, detection limits when the marsh was flooded. Inorganic N concentrations increased markedly in the late dry season as water levels receded in the marsh. These increases were most likely due to decomposition of organic matter and release of ammonia and nitrates into the shallow water column.

In contrast, total organic nitrogen concentrations at both background and peripheral marsh sites were comparable to eutrophic Florida lakes with background sites averaging between 2.04-2.90 mg/l (Table 3). The origin of these higher total organic N levels is not well understood; however, possible sources include rainfall, decomposition of leaf litter or peat soils, and leaching from living plants. Data collected from south Florida agricultural sites show significant amounts of total N present in rainfall ranging from 0.6-1.1 mg/l and undoubtedly represent a major source of N to interior marsh sites (Shannon, 1978, see also Table 2). High total N concentrations within marsh surface waters commonly occur in temperate and south Florida wetlands (Davis, 1981 and references cited therein).

Total dissolved PO₄ concentrations at interior marsh sites showed little change over time, remaining near detection levels during high and low water periods. Phosphorus concentrations were highest just prior to marsh drying, and after reflooding due to the mineralization of organic matter. However, these higher concentrations were of short duration, and returned to background levels within a month after the sites were flooded (Figures 10-13). Similiar trends of low nutrient availability were also encountered at the 1-8T gauge located on the eastern peripheral marsh of WCA-1 (Figure 14).

In contrast, peripheral marsh sites (B-2 and B-5) located near the S-10 discharge structure in WCA-2A showed a positive correlation with high water levels, phosphorus, and chloride content (R=.690). Concentrations of dissolved nutrients were highest at

TABLE 3. A COMPARISON OF AVERAGE NUTRIENT CONCENTRATIONS AT INTERIOR WCA MARSH SITES WITH FLORIDA LAKES AND SELECTED LAKE TROPHIC STATE CLASSIFICATION SCHEMES										
INTERIOR MAI	Total P (mg/l) RSH SITES	Total N (mg/l)	Total Org-N (mg/l)	Inorganic N (mg/l)						
WCA-1	.014 (.024)	2.86 (3.39)	2.72 (2.89)	.09 (.29)						
WCA-2A	.011 (.023)	3.05 (1.91)	2. 9 0 (1.70)	.12 (.43)						
WCA-3A	.009 (.017)	2.15 (1.27)	2.04 (1.11)	.09 (.28)						
"CLEAR" NOR (Brezonik and	-									
Oligotrophic	.013 (.003)	*	.25 (.08)							
Mesotrophic	.023 (.014)		.75 (.30)							
Eutrophic	.306 (.251)		1. 98 (1.10)							
GENERALIZED (Likens, 1975)	LAKE TROPHI	IC STATE CLA	ASSIFICATIO	N						
Oligotrophic	<.001010	<.00160								
Mesotrophic	.010030	.50-1.10								
Eutrophic	>.300	>1.00								
EUROPEAN LA (Vollenweider		STATE CLASS	SIFICATION							
Ultra-oligotro	ohic .005			<.20						
Oligotrophic	.005010			.2040						
Mesotrophic	.010030			.3065						
Eutrophic	.030100			.50-1.50						
Hypereutroph	ic >.100			>1.5						
Upper value re	presents mear	n, lower value	e in parenth	eses = standard						

peripheral marsh sites which received nutrient enriched canal water inflows from the three S-10 structures (Figures 15 and 16). Site B-2, located 0.6 km south of S-10C in WCA-2A, exhibited consistently high concentrations of both inorganic N and total dissolved PO₄ from 1978-1981 (B-2 was not monitored during 1982). During 1978-1980, Site B-5 (located 4.2 dominant species in most natural substrate collections

*---Parameter not reported

substrate collections (i.e., submerged periphyton mats, km south of S-10C) had N and P concentrations that were at or near background levels. Nutrient concentrations at Site B-5 increased dramatically following reflooding of the marsh after the 1981 drought (Figure 16). These data indicate that nutrients have penetrated the WCA-2A marsh much further than previously reported in our 1978-79 study (Swift, 1981).

Periphyton Species Composition

Water Conservation Area periphyton communities grown on glass slides contained a total of 333 taxa (including species and varieties) of epiphytic algae representing 107 genera and eight divisions (Appendix A, Table A-1).

1. Myxophyceae (Blue-greens). In terms of cell volume, biomass (as measured by chlorophyll a) and areal coverage, filamentous blue-greens were the most abundant algal group colonizing both glass slides and natural substrates in WCA-2A, 3A, and the peripheral marsh of WCA-1. These blue-green communities closely resemble the "calcareous periphyton" flora described by Van Meter (1965, 1973) and Gleason and Spackman (1974) as the dominant algae of the Everglades region. Of the fifteen bluegreen taxa identified, only three were numerically important in terms of cell volume: Schizothrix calcicola (Ag.) Gom., Scytonema hofmannii Ag., and Microcoleus lyngbyaceus (Kutz) Crouan.

The most widely distributed and most frequently encountered alga observed in the study was <u>Schizothrix calcicola</u>, occuring in every sample collected. This blue-green represented the bulk of periphyton biomass grown on glass slides in shallow, low nutrient, calcium rich waters, and was also a

dominant species in most natural substrate collections., (i.e., submerged periphyton mats, and as epiphytes of <u>Utricularia spp.</u>, <u>Eleocharis spp.</u> and sawgrass stems, etc.). Appendix B "Taxonomic Notes" provides a more detailed description of the morphological variations associated with the identification of <u>S. calcicola</u> from our collections.

The second most abundant filamentous blue-green was Scytonema hofmannii, occurring at times as a co-dominant species in association with Schizothrix calcicola at calcium rich interior marsh sites. Scytonema hofmannii was a relatively slow growing species and was not as commonly encountered in our glass slide collections as compared to periphyton mat cores collected from marsh sediments. This filamentous blue-green accounted for a maximum of 45 percent of the total periphyton community (by volume) on glass slide substrates whereas S. hofmannii accounted for 50-80 percent of the algae collected from periphyton mat cores. Scytonema hofmannii and Schizothrix calcicola were the dominant periphyton flora in waters containing high concentrations of dissolved minerals (bicarbonate and calcium) and low concentrations of inorganic nutrients.

The filamentous blue-green, Microcoleus lyngbyaceus, was the most common algae reported from nutrient enriched peripheral marsh sites. In the presence of high concentrations of inorganic nitrogen and phosphorus, this species was responsible for some of the largest biomass increases reported in this study.

Other common (but rarely abundant) filamentous blue-green species included Spirulina subsalsa, Johannesbaptista pellucida, Oscillatoria lutea, Oscillatoria cf. tenius, and Stigomena sp.

Coccoid blue-greens were not important components of the WCA periphyton and usually represented less than one percent of the community by volume. The most common coccoid blue-greens present were <u>Anacystis dimidiata</u>, <u>Anacystis montana</u>, and Coccochloris elabens.

2. <u>Bacillariophyceae (Diatoms)</u>. Diatoms were the second most important group of algae colonizing glass slides. Diatoms included 134 species represented by 25 genera (Table A-1, Appendix). Diatom communities from hardwater (calcium rich), low nutrient, interior marsh sites were characterized by the following species: <u>Anomoeneis vitrea</u>, <u>Cymbella ruttneri</u>, <u>Mastogloia smithii v. lacustris</u>, <u>Cymbella minuta v. psuedogracilis</u>, <u>Synedra pahokeensis</u> sp. nov., <u>Gomphonema</u> ef. <u>affine v. insigne</u>, <u>Nitzschia storchii sp. nov.</u>, and <u>Amphora venta</u>.

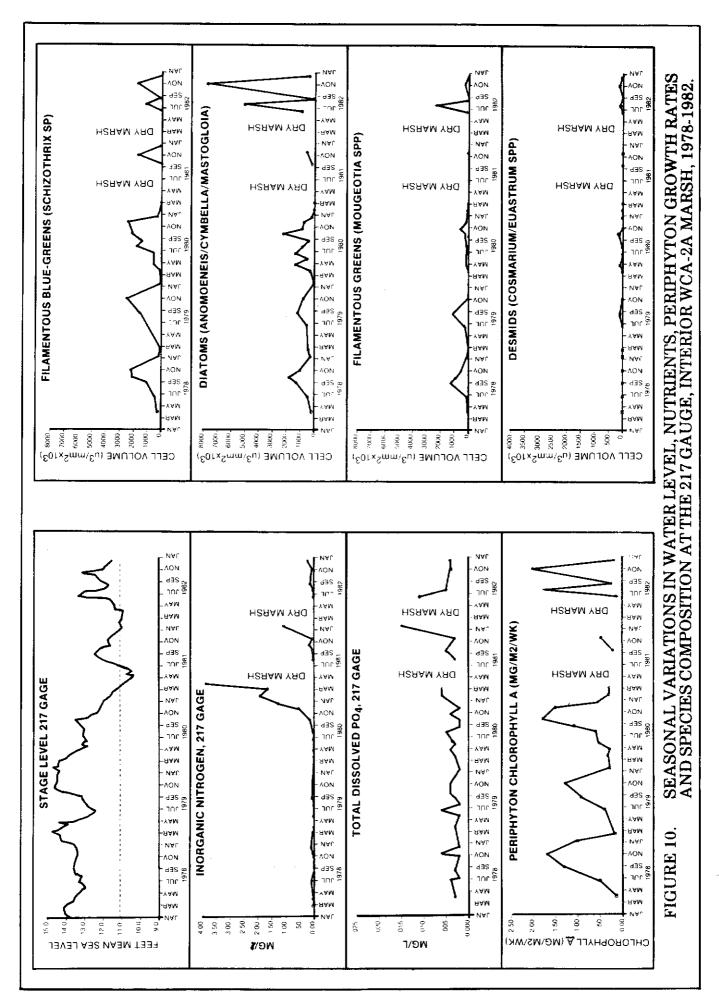
TABLE 4. **AVERAGE NUTRIENT CONCENTRATIONS AT INTERIOR** WCA SITES, FEBRUARY 1977 - JUNE 1983 Element All Sites Combined WCA-1 WCA-2A WCA-3A (mq/l)Inorganic N .09 .10 .09 .12 (as NO₃-N, (.29)(.43)(.28)(.33) NO_2-N , NH_4-N) Total N 3.05 (TKN + 2.53 2.86 2.15 (3.39)(1.91)(1.27)Inorganic N) (2.35)Total 2.41 2.90 2.04 Organic N 1.71 (1.70)(1.11)(1.91)(2.89)(TKN-NH₄) .009 Total PO₄ .011 .014 .011 (.021)(.024)(.023)(.017)Total .006 .005 .005 .006 Dissolved PO₄ (.006)(.007)(.004)(.011)Ortho-PO₄* .003 .003 <.002 .004 (.001)(.007)(.004)(.002)Number of Analysis 390 121 125 117

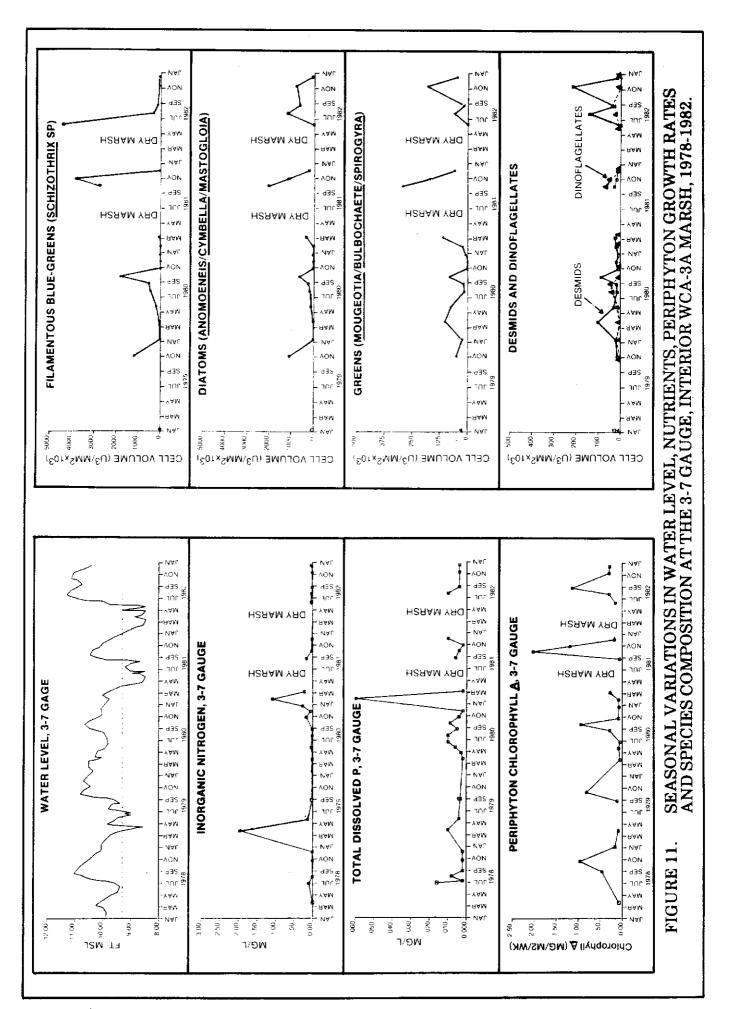
Upper value represents mean, lower value in parentheses is standard deviation

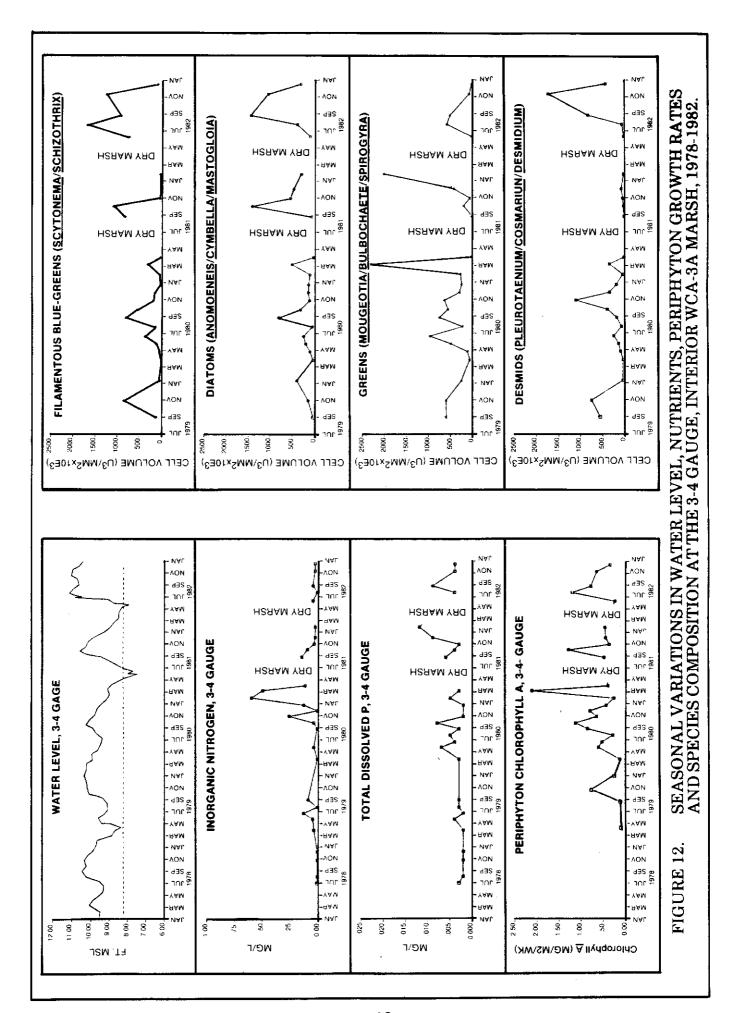
The most unique diatom assemblages occurred in the interior WCA-1 marsh (Site A-3). Surface waters at Site A-3 were acidic (pH 5-6) and low in ionic content (soft water) with moderately low concentrations of nutrients present. Characteristic diatoms of the interior of WCA-1 were Cymbella amphioxys, Anomoeoneis serians v. brachysira, Anomoeoneis serians, Anomoeoneis vitrea, Frustulia rhomboides v. saxonica, F. rhomboides v. crassinervia, Cymbella minuta v. silesiaca, Nitzschia sp. 7 sp. nov., Eunotia naegelii, Synedra tenera, Pinnularia biceps, Navicula subtilissima, and Stenopterobia intermedia.

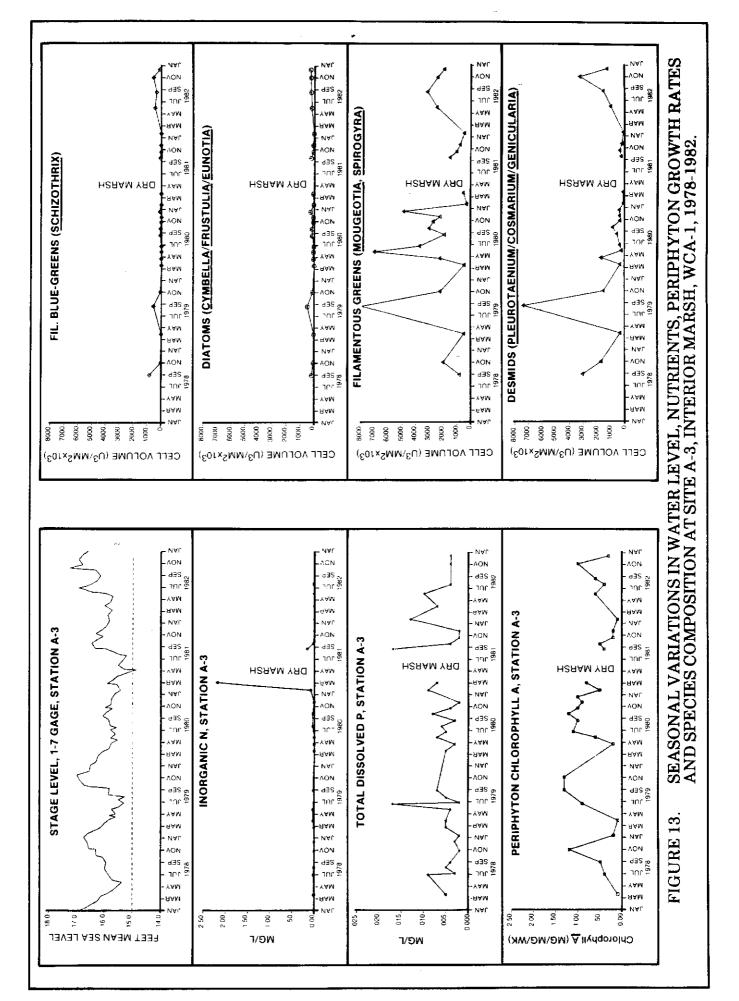
Peripheral marsh sites influenced by nutrient enriched canal water also supported a unique diatom flora. Common diatoms occurring at nutrient enriched marsh sites were <u>Nitzschia amphibia</u>, <u>Gomphonema parvulum</u>, <u>Nitzschia palea</u>, <u>Nitzschia tarda</u>, <u>Nitzschia sigmoidea</u>, <u>Nitzschia sp. 7</u>, <u>Navicula confervacea</u>, <u>Navicula disputans</u>, and <u>Navicula cuspidata</u>.

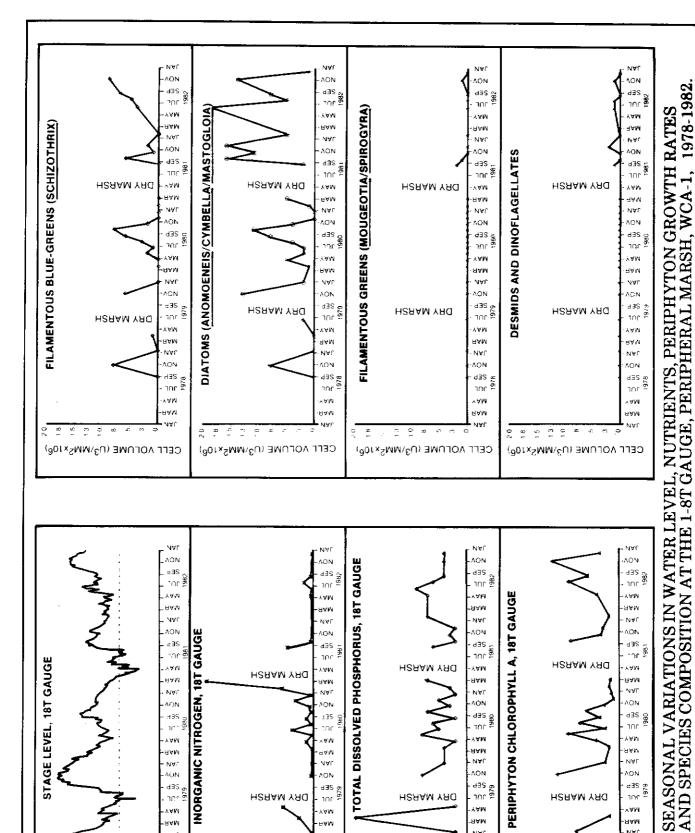
^{*}Average ortho-PO₄ is an artificially high value. Limits of detection were changed from less than .002 mg/l to less than .004 mg/l midway through the study. Average ortho-PO₄ values are probably less than .002 mg/l at most interior sites.

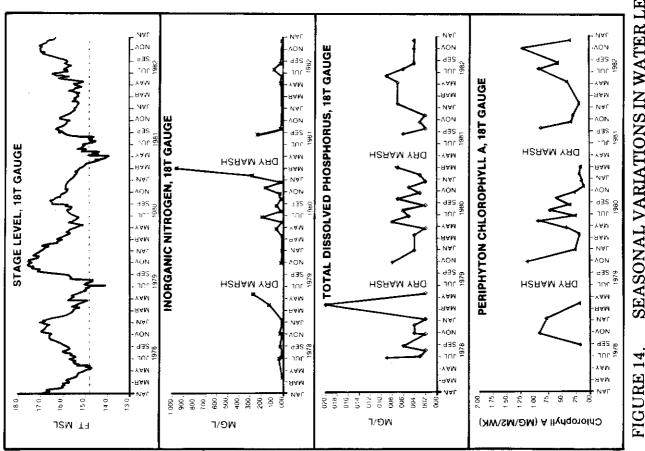


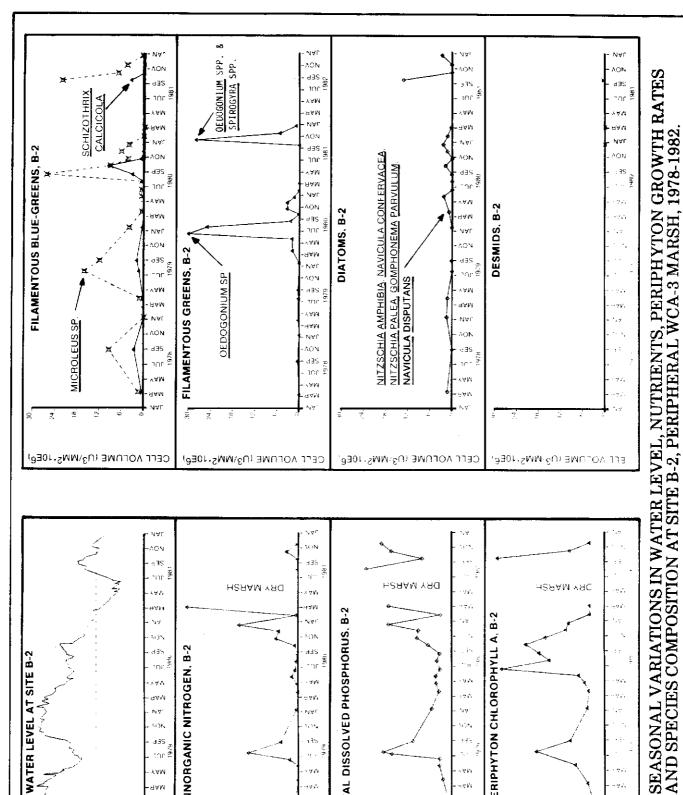












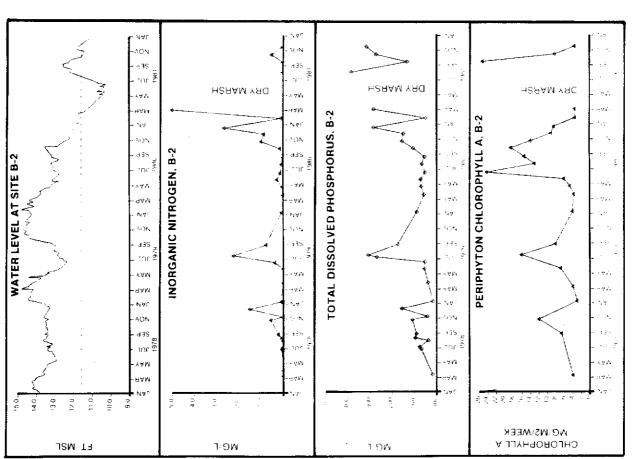
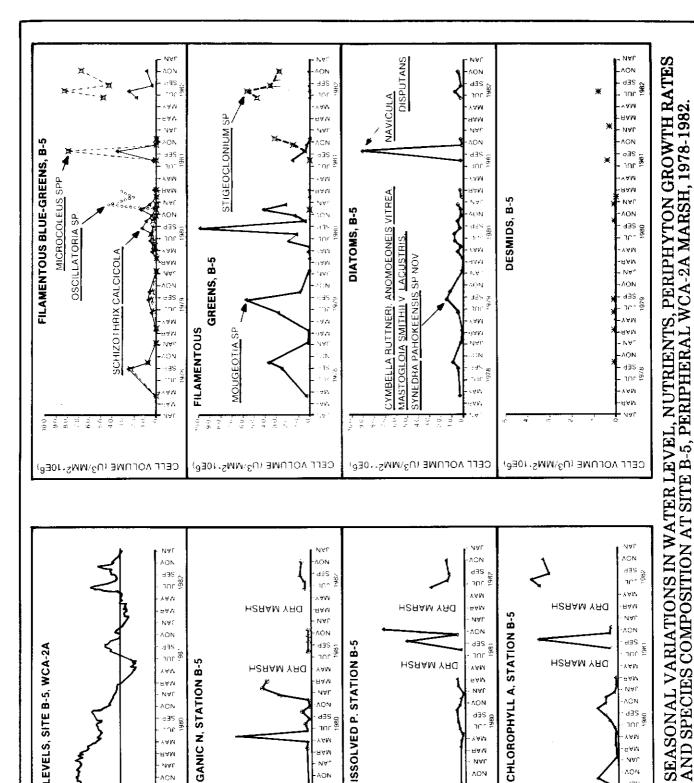


FIGURE 15.



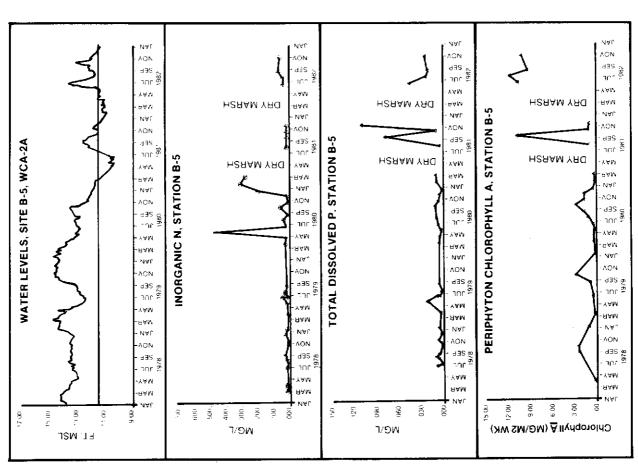


FIGURE 16.

Four diatom species recorded in this study represent new species to the literature (pers. comm. Dr. Charles Reimer, Academy of Natural Sciences of Philadelphia). Tentative names given to these new species are: Synedra pahokeensis sp. nov., Synedra sp. 2, Nitzschia storchii sp. nov., and Nitzschia sp. no. 7.

3. Chlorophyceae (Green Algae). Filamentous green algae and desmids were the two most important chlorophycean groups present. The five most important genera of filamentous greens were Mougeotia, Spirogyra, Bulbochaete, Oedogonium, and Stigeoclonium. Mougeotia was the most common and widely distributed of these genera.

Two families, Mesotaeniaceae and Desmidiaceae (collectively known as desmids), were important members of the periphyton community in the interior marsh of WCA-1 and WCA-3A. The acid, soft water marsh of interior WCA-1 supported a highly diverse periphyton community of desmids and filamentous green algae. The desmids included 122 species recorded from Site A-3 (Table 5). The six most common desmid genera were as follows (listed in descending order of importance): Pleurotaenium, Cosmarium, Genicularia, Desmidium, Euastrum, and Other common genera included Staurastrum. Arthrodesmus, Bambusina, Closterium, Cylindrocystis, Gonatozygon, Hyalotheca, Micrasterias, Netrium, Sphaherozosma, Spondylosium, Triploceras, and Xanthidium.

- 4. <u>Dinophyceae</u>. Although dinoflagellates are most often associated with marine plankton, two motile, "armored" (cells covered with interlocking plates) fresh water species, <u>Peridinium inconspicum</u> and <u>Peridinium</u> sp. 2, were common components of the WCA periphyton. Dinoflagellates were never present as dominant or co-dominant species but were intermittantly observed as rare species.
- 5. Chrysophyceae. The majority of chrysophytes identified were epiphytes on filamentous green algae. Although this group includes a large variety of plant forms, the most commonly encountered chrysophyte consisted of a naked cell enclosed within a lorica or "shell". Chrysophytes were widely distributed throughout the WCA, but represented only a minor fraction of the periphyton community (usually less than 1%). Six genera were recorded: Derepyxis, Dinobryon, Epipyxis, Lagynion, Mallomonas, and Synura.
- 6. <u>Euglenophyceae</u>, 7. <u>Cryptophyceae</u>, and 8. <u>Charophyceae</u>. These three remaining divisions were not important components of the WCA periphyton community and represented only a minor fraction of the periphyton assemblage (Table A-1).

Geographical and Seasonal Distribution of Major Algal Groups

Comparisons of the spatial and seasonal distributions of the dominant taxa were made between interior and peripheral marsh sites and among the three Water Conservation Areas.

Water Conservation Area 2A. One peripheral site (B-2), one transitional site (B-5), and one interior marsh site (217 gauge) were monitored. Sites B-5 and the 217 gauge were monitored from 1978-1982; site B-2 was sampled from 1978-1981.

Interior marsh (217 gauge): In terms of cell volume and chlorophyll a content, filamentous blue-greens (Schizothrix calcicola) and an assemblage of hardwater diatoms (Anomoeneis vitrea, Mastogloia smithii v. lacustris, Cymbella ruttnerii, and Synedra pahokeensis sp. nov.) represented the dominant periphyton flora at the interior 217 gauge site from March 1978-March 1981. Filamentous blue-greens and diatoms were most abundant during September and October (end of wet season) and least abundant during January and February (Figure 10). After reflooding of the marsh in June 1982, diatom populations at the 217 gauge increased rapidly and replaced filamentous blue-greens as the dominant flora. Filamentous greens (primarily Mougeotia sp.) were the third major group present. Peak abundance of Mougeotia occurred in September and October of each year, with the exception of 1982 when largest populations were recorded in July (Figure 10). Total numbers of algal species present at the 217 gauge were low in comparison to other interior marsh sites and included only blue-greens, greens, and diatoms. Desmids, dinoflagellates, and chrysophytes were absent or only present as rare components of the 217 gauge microflora.

Peripheral and transitional marsh sites B-2 and B-5: Periphyton species composition and biomass at these two sites were markedly different from those recorded at the 217 gauge. Site B-2, located 0.6 km south of S-10C, was dominated by the filamentous blue-green Microcoleus lyngbyaceus in association with an assemblage of diatoms and two genera of filamentous green algae (Oedogonium and Spirogyra). Seasonal maximum abundance of Microcoleus occurred in the late summer months (August-September) during periods of high water levels and high concentrations of dissolved nutrients (Figure 15). Lowest populations generally occurred during January and February.

Filamentous green algae (Oedogonium spp. and Spirogyra spp.) showed no clear seasonal trends at Site B-2. However, these two genera represented the dominant flora and the highest cell volumes recorded (51 x 106 microns³/mm²) during June and July of 1980

(Figure 15). Diatom communities at Site B-2 also showed no clear seasonal pattern. The majority of diatom species recorded at Site B-2 were species that are most often associated with nutrient enriched waters (Lowe, 1974). The most common diatom species at Site B-2 included Nitzschia amphibia, Nitzschia palea, Gomphonema parvulum, Navicula confervacea, and Navicula disputans.

From 1978-1980, two filamentous greens (Mougeotia spp. and Spirogyra spp.) were the dominant algae at Site B-5 located 4.2 km south of the S-10C structure. Filamentous greens were most abundant during the late summer months, while sparse populations of filamentous blue-greens and diatoms dominated this site during winter months (Figure 16).

After the 1981 drought and subsequent reflooding of the marsh via discharges from the S-10 structures, Site B-5 experienced major changes in periphyton species composition in response to increases in the phosphorus content of marsh surface waters. From 1981-1982, Site B-5 was dominated by heavy growths of the filamentous blue-green (Microcoleus lyngbyaceus), the filamentous green (Stigeoclonium spp.), and the pollution indicator diatom Navicula disputans (Figure 16).

Water Conservation Area 3A. Two interior marsh sites (3-4 and 3-7 gauges) were sampled from 1978-1982. Filamentous blue-greens (Schizothrix calcicola and Scytonema hofmannii) were the dominant algae at the 3-7 gauge (Figure 11), while filamentous blue-greens and diatoms (Anomoeneis vitrea, Cymbella ruttneri, Mastogloia smithii v. lacustris, and Synedra pahokeensis sp. nov.) were the two most abundant periphyton groups at the 3-4 gauge (Figure 12). Filamentous green algae (Mougeotia, Bulbochaetae, and Spirogyra) and desmids (Pleurotaenium spp. and Cosmarium spp.) were also important seasonal components of the 3-4 gauge periphyton.

Maximum abundance of Schizothrix and Scytonema occurred from late August through October of each year at both the 3-4 and 3-7 gauges with lowest populations present in January and February. Maximum diatom abundance generally coincided with maximum filamentous blue-green abundance. Filamentous greens (Mougeotia sp.) at the 3-4 gauge reached peak abundance during low water conditions (just before the marsh dried) and suggests a response to increased inorganic nitrogen levels. Desmids reached peak abundance at the 3-4 gauge during August-October in the years 1979, 1980, and 1982. During 1981, desmids were not important components of the 3-4 gauge periphyton. Populations of desmids and filamentous green algae at the 3-7 gauge remained low throughout the study.

<u>Water Conservation Area 1</u>. One peripheral and one interior marsh site were monitored from 1978-1982 (1-8T gauge and Site A-3).

Peripheral marsh (1-8T gauge): Water quality at the 1-8T gauge was high in dissolved minerals due to its close proximity to the L-40 Canal (0.6 km) but had low concentrations of nutrients (comparable to sites 3-4 and 3-7 in WCA-3A). Diatoms dominated the periphyton community at the 1-8T gauge site with filamentous blue-greens (Schizothrix calcicola) representing the second most important group (Figure 14). The most common diatom species encountered at the 1-8T gauge were Mastogloia smithii v. lacustris, Anomoeoneis vitrea, Cymbella ruttneri, Synedra pahokeensis sp. nov., and Cymbella minuta v. psuedogracilis. Seasonally, peak abundance of both diatoms and S. calcicola occurred in September of each year in association with rising water levels (Figure 15). Minimum abundance occurred during January through March. The maximum standing crop of diatoms occurred in September 1982 when the marsh was reflooded after an extended drought. Filamentous greens, desmids, and dinoflagellates were not important components of the periphyton at the 1-8T gauge.

Interior marsh (site A-3): Periphyton communities collected from the acid, soft water interior marsh of WCA-1 (Site A-3) represented the most unique and diverse algal flora encountered in the study. Algal communities at Site A-3 were dominated by the Chlorophyceae (green algae) represented by two major groups, filamentous greens (Mougeotia spp. and Spirogyra spp.) and desmids. Filamentous greens reached maximum abundance during June-August of 1979, 1980, and 1982, with lowest populations occurring during winter months. Filamentous greens were also low in abundance after reflooding of the marsh in 1981 (Figure 13). Desmids were most common during the months of August 1978 and 1979 and September 1982. The desmid population remained low throughout 1980 and 1981 (Figure 13). The most common desmid genera encountered were Pleurotaenium, Cosmarium, Genicularia, and Desmidium. Populations of filamentous blue greens and diatoms remained low at Site A-3 throughout the four years of study. Site A-3 supported an association of diatom species which have been reported as reliable indicators of acid, softwater environments (Patrick and Reimer, 1966; Round, 1965; Lowe, 1974; Patrick, 1977).

Periphyton Growth Rates

Active chlorophyll <u>a</u> extracted from algal pigments and corrected for phaeophytin degradation products (Strickland and Parsons, 1968) was used as an indicator of periphyton growth rates or biomass accumulation over a specified time period. Periphyton

growth rates are reported as mg chlorophyll <u>a</u> per square meter per week (mg chl <u>a</u>.m- 2 .wk- 1).

Periphyton growth rates at interior marsh sites ranged from 0.1-2.3 mg chl a.m-2.wk-1 and were low in comparison to nutrient enriched peripheral marsh sites (Figures 10-16). Maximum growth rates at interior marsh sites occurred during September and early October (end of wet season). Maximum growth rates at interior sites in WCA-2A and WCA-3A and the peripheral marsh of WCA-1 were primarily due to the abundance of filamentous blue-greens (Schizothrix calcicola and Scytonema hofmanii) and an associated diatom flora. In the interior marsh of WCA-1, filamentous greens (Mougeotia spp. and Spirogyra spp.) and desmids were responsible for peak chlorophyll a concentrations.

Periphyton growth rates at nutrient enriched marsh sites B-2 and B-5 were more than 10 times those recorded from interior marsh sites (Figures 15 and 16). Periphyton growth rates at Site B-2, located 0.6 km south of S-10C, were consistently high throughout the study, reaching a maximum rate of 25.0 mg chl a·m·2·wk·1 during June 1980 and September 1981. Peak growth rates during June 1980 were due to the abundance of Oedogonium spp., while Microcoleus lyngbyaceus dominated Site B-2 during the fall of each year.

Seasonal periphyton growth rates at interior marsh sites were relatively independent of surface water levels and nutrient concentrations. On an annual basis, interior marsh periphyton growth rates responded more closely to seasonal variations in light and water temperature than to variations in nutrients or water depth (see Figure 3). Chlorophyll a levels were generally lowest during January-March. These low values corresponded with low seasonal water temperature and light levels experienced each winter. As water temperatures and light levels increased during April-June there was a corresponding increase in periphyton growth rates. Growth rates continued beyond the period of maximum light and temperature so that peak seasonal concentrations of chlorophyll a occurred during September and early October of each year. These values occurred approximately four to six weeks out of phase, with maximum water temperature and two months lag time behind maximum solar radiation levels.

Periphyton Nutrient Content

Gerloff and Skoog (1954) demonstrated that concentrations of nitrogen and phosphorus in algal cell tissue can be used to measure the availability of these elements in surface waters.

The phosphorus content of periphyton communities collected from interior marsh sites in

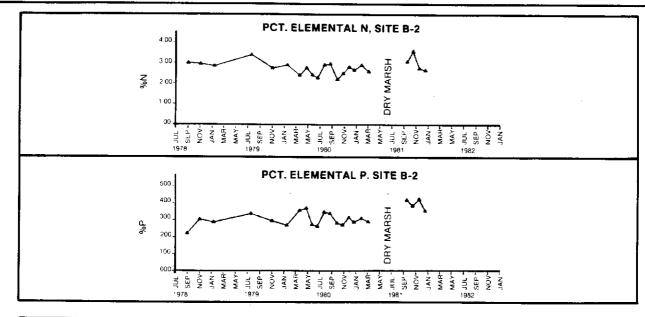
WCA-2A and WCA-3A were very low (0.01-0.04% P) in comparison to samples collected from the interior of WCA-1 and nutrient enriched peripheral marsh sites (Figures 17 and 18 a, b). Highest elemental P concentrations were observed at sites B-2 and B-5. From 1978-1980, phosphorus levels in the water at B-5 were very low, averaging about 7 µg P/I. Algae growth rates were also low and had low concentrations of elemental P in cell tissues. After the 1981 drought, increased sheet flow across WCA-2A, as a result of heavy autumn rains and high discharges through the S-10 structures, caused nutrient enriched canal water to penetrate the marsh much further than previously Consequently, marsh water nutrient content, periphyton growth rates, and elemental P content of algae at site B-5 showed dramatic increases during 1981 and 1982 (Figure 17).

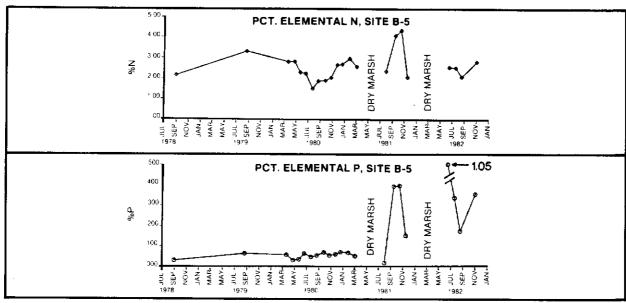
Periphyton elemental P concentrations at Site A-3 (WCA-1) ranged from .048 - 0.094 percent. Periphyton communities in the interior of WCA-1 were dominated by filamentous green algae and desmids and contained roughly 2-3 times as much phosphorus as the blue-green dominated periphyton communities of the interior of WCA-2A and WCA-3A (Figure 18a).

Seasonal variations in the elemental P content of interior marsh periphyton were not distinct. However, maximum periphyton phosphorus content generally coincided with maximum periphyton growth rates which occurred during late summer and early fall (end of wet season).

In WCA-2A and WCA-3A linear correlations between the elemental N content of the algae and marsh water inorganic nitrogen concentrations were not readily apparent. Periphyton elemental N levels were generally highest during August-early October coinciding with maximum algal growth rates (Figures 17, 18a, 18b). However, there were several exceptions. After reflooding of the marsh in 1981, periphyton elemental N concentrations at Site B-5 rapidly increased in response to elevated surface water nutrient concentrations. During March 1981, elemental N concentrations at sites A-3 and the 1-8T gauge (WCA-1) increased rapidly in response to high levels of inorganic nitrogen during low water conditions (Figure 18a).

Comparisons of average periphyton nitrogen to phosphorus mass (N:P) ratios among the seven sites are presented in Figure 19. High N:P ratios were typical of interior marsh sites in WCA-2A, WCA-3A, and the peripheral marsh of WCA-1 with average mass ratios ranging from 101:1-107:1. Lowest N:P ratios (average 8.6:1) were encountered at the peripheral marsh site (B-2) located south of S-10C in WCA-2A. Nitrogen:phosphorus mass ratios at Site A-3 (avg. = 51.5:1) located within the interior of





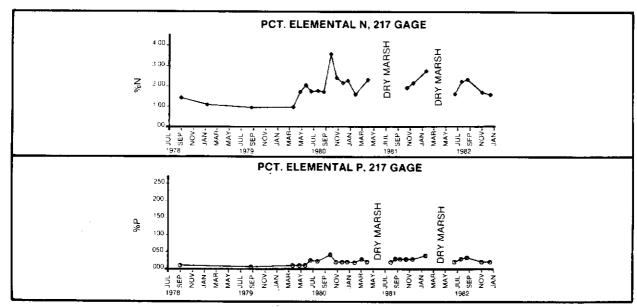
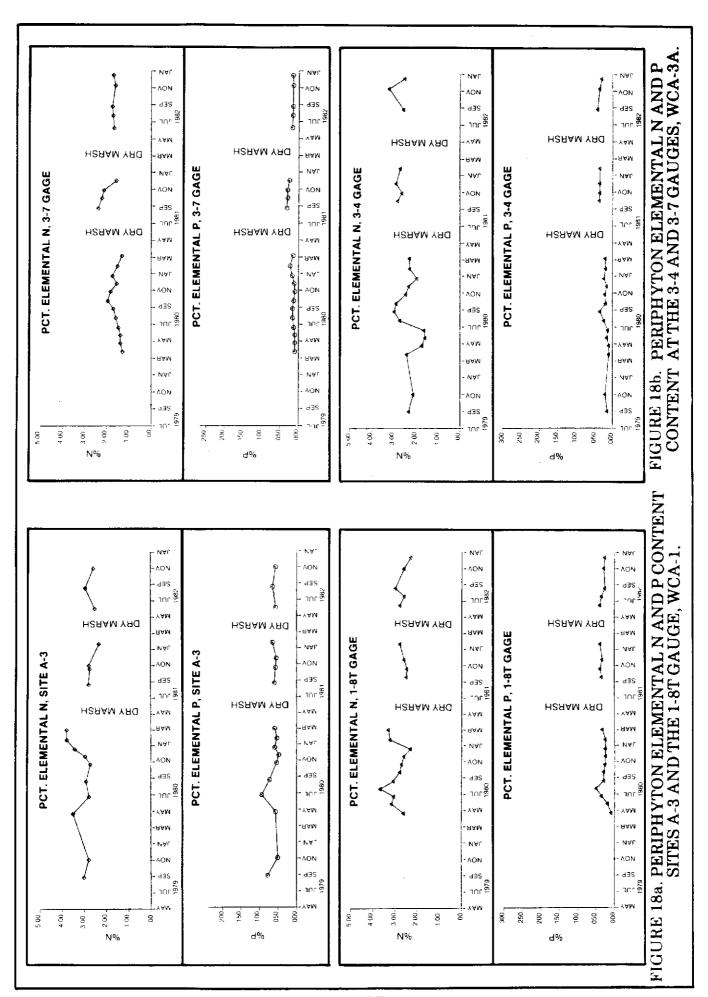


FIGURE 17. PERIPHYTON ELEMENTAL N AND P CONTENT AT SITES B-2, B-5 AND THE 217 GAUGE, WCA-2A.



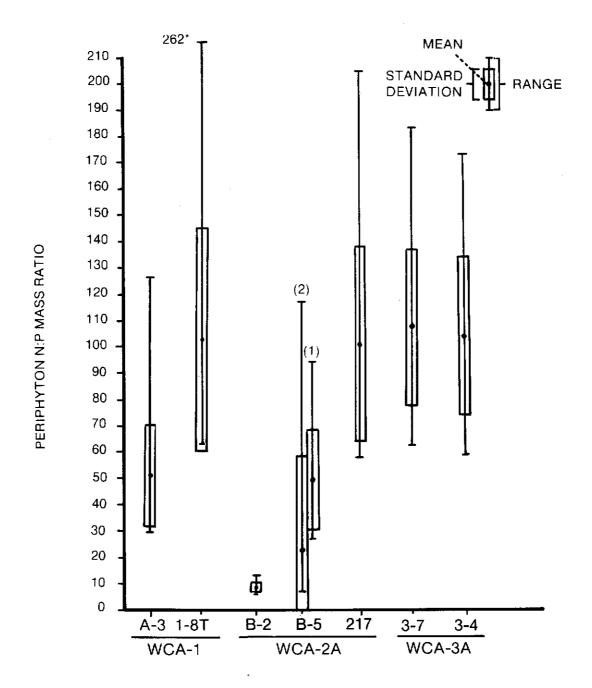


FIGURE 19. PERIPHYTON N:P RATIO AVERAGES, STANDARD DEVIATIONS AND RANGES AT SEVEN LONG TERM WCA MONITORING SITES.

(1) AVERAGE N:P RATIO PRIOR TO 1981 DROUGHT (1978-MARCH 1981)

(2) AVERAGE N:P RATIO AFTER 1981 DROUGHT AND REFLOODING OF MARSH JULY 1981 - DECEMBER 1982 (showing impact of nutrients on Site B-5)

WCA-1 were generally lower than interior WCA-2A and WCA-3A sites.

Marsh water phosphorus showed a significant inverse correlation with the N:P content of the periphyton. As the phosphorus content of the marsh water increased, algae N:P mass ratios decreased exponentially (Figure A-1, Appendix A). A similar relationship also existed between periphyton growth rates and the N:P content of the algae. As chlorophyll a concentrations increased, algae N:P mass ratios decreased logarithmically. Maximum algae growth rates in the Water Conservation Areas occur at N:P mass ratios of less than 15:1 (Figure A-2, Appendix A).

Species Diversity

Periphyton species diversity at Site A-3 was significantly higher (Tukey's test, p < .05) than all other sites sampled (Figure 20). This was a result of the rich and varied desmid flora (i.e., rich in terms of total number of species) that were present at Site A-3 throughout the study. High species diversity, and high relative abundance of desmids, were closely linked to the mineral content of marsh surface waters (Tables 5 and 6). An inverse relationship exists between periphyton species diversity and calcium carbonate alkalinity of WCA marsh waters (Figure A-3, Appendix A). Highest species diversity and high desmid relative abundance occurred primarily in soft water, with lower diversity indices occurring in hard water.

Lowest species diversity indices were recorded at nutrient enriched Site B-2 as a result of the dominance of <u>Microcoleus lyngbyaceus</u> and <u>Oedogonium</u> spp. The filamentous blue-green <u>M. lyngbyaceus</u> reached maximum abundance in the late summer and fall, while the filamentous green, <u>Oedogonium</u> spp., dominated Site B-2 during the early summer months (Figure 15). No significant differences in algal species diversity were noted among the remaining five sites (i.e., Sites B-5, 1-8T, 217, 3-4, and 3-7 gauges).

Relationship Between Periphyton Species Composition and Environmental Factors

Factor analysis reduced 30 variables to 5 biologically interpretable factors that accounted for 74% of the variance within the data set (Table 5). The primary factor (Factor 1) was closely related to the following variables: chlorides, conductivity, hardness, silicon, alkalinity, Microcoleus relative abundance, and (-) desmid relative abundance. Factor 1 might be labeled "periphyton response to surface water major ion concentration." Factor 1 shows that waters containing low concentrations of dissolved minerals were highly correlated with the development of a desmid dominated periphyton community; while marsh waters containing high concentrations of major

ions favored periphyton communities that were dominated by the filamentous blue-green, <u>Microcoleus</u> spp.

Factor 2 shows a significant correlation between periphyton species diversity and the occurrence of filamentous greens, desmids, dinoflagellates, and a group of diatom species indicative of acid, soft water conditions (i.e., Cymbella amphioxys, Frustulia rhomboides v. saxonica, Anomoeoneis serians v. brachysira, etc.). These correlations are a result of the unique and varied periphyton assemblage that occurs in the acid, soft water interior marsh of WCA-1. The desmid flora of the interior of WCA-1 was exceedingly rich and contained over 120 species (Table A-1). As a result, desmids were significantly correlated with high species diversity indices. Desmids were typically found in close association with filamentous greens (Mougeotia and Oedogonium), acid water diatoms, and dinoflagellates (Peridinium spp.). As a result, these algae groups were also correlated with high species diversity.

Factor 3 was related to the following variables: Schizothrix calcicola, and Scytonema hofmannii relative abundance, "alkaline" water diatoms (i.e. Mastogloia smithii v. lacustris, Cymbella ruttneri, Synedra pahokeensis sp. nov., Anomoeoneis vitrea) and the calculated mass ratio of elemental N:P within periphyton cell tissue. Factor 3 shows that Schizothrix-Scytonema- alkaline water diatom communities reach maximum abundance under conditions of low phosphorus supply (which results in a high N:P ratio). These results suggest these algal species may be used as biological markers of calcium rich, low nutrient, water quality conditions in the Water Conservation Areas.

Factor 4 is related to <u>Stigeoclonium</u> relative abundance, eutrophic indicator diatoms (i.e., <u>Nitzschia amphibia</u>, <u>Navicula disputans</u>, <u>Navicula confervaceae</u>, <u>Gomphonema parvulum</u>, <u>Nitzschia palea</u>) relative abundance, marsh water total dissolved PO₄ concentration, and periphyton elemental P content. The relationship within factor 4 shows that <u>Stigeoclonium</u> and eutrophic diatoms reach maximum abundance under phosphorus enriched conditions and suggests that these algae may serve as useful indicator species of nutrient enrichment.

The final factor (Factor 5) is related to water depth, periphyton elemental P content, Microcoleus relative abundance, periphyton growth rates, and marsh water total dissolved PO₄ content. Factor 5 relates primarily to the deep water, nutrient enriched environment that was prevalent at Sites B-2 and B-5. These sites were unique because they produced the highest periphyton biomass (as chlorophyll <u>a</u>) of all sites studied and were seasonally dominated by the

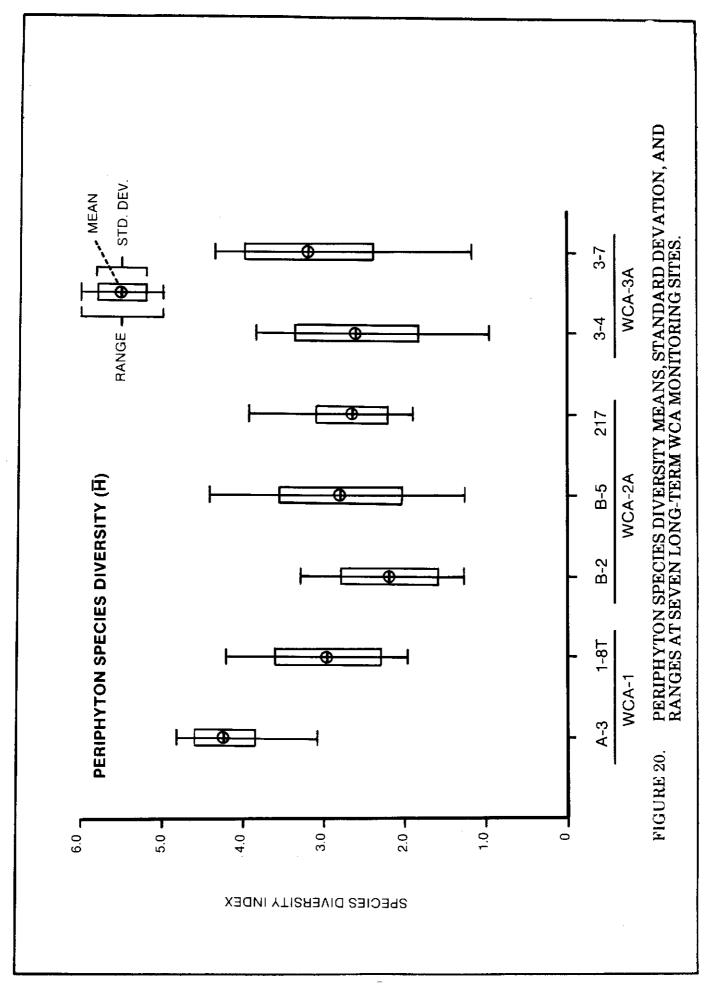


TABLE 5.	VARIMAX ROTATED FACTOR LOADINGS
	FOR 30 BIOLOGICAL AND PHYSIO-
	CHEMICAL VARIABLES,* 1978-1982

Variable	R +
FACTOR 1	
CL**	.927
Conductivity	.919
Hardness	.901
SiO ₂	.753
Alkalinity	.708
% Microcoleus	.497
% Desmids	478
Variable	R
FACTOR 2	
Species Diversity(H)	926
% Mougeotia/Oedogonium	684
% Desmids	.630
% Acid Water Diatoms	.472
% Dinoflagelletes	.431
FACTOR 3	
% Schizothrix calcicola/	
Scytonema hoffmanni	.939
% Alkaline Water Diatoms	.894
Periphyton Cell N:P Ratio	.847
FACTOR 4	
% <u>Stigeoclonium</u>	.762
%. Eutrophic Indicator Diatoms	.733
Total Dissolved PO ₄	.721
Periphyton Elemental P Content	.450
FACTOR 5	
Water Depth	.819
Periphyton Elemental P Content	.777
% Microcoleus	.560
Periphyton Chlorophyll <u>a</u>	.542
Total Dissolved PO ₄	.483
l .	

Legend:

- Of the 30 variables tested, only % cell N content was not significantly correlated with another parameter.
- ** Because of the high degree of correlation between a number of major ion species and chlorides (r = .90) the following ions were omitted from the above analysis to overcome problems of multicollinearity. These ions were: Na, K, Mg, and Ca.
- + Coefficient (R) values represent both regression weights and correlation coefficients (Nie et. al. 1970, SPSS).
- n = 198 observations, significant values of R at 5% level = 0.431 at 1% level = .460.

filamentous blue-green <u>Microcoleus</u> spp. These relationships suggest that <u>Microcoleus</u> may also be a useful marker species of phosphorus enrichment in the Water Conservation Areas.

In summary, factor analysis shows the importance of major ions, phosphorus concentrations, and water depth on periphyton species composition, cell nutrition and algal growth rates in the Water Conservation Areas.

Stepwise multiple regression analyses closely paralleled results of factor analysis (Table 6). Marsh water phosphorus content and water depth were highly correlated with increases in periphyton growth rates (as measured by chlorophyll a) and increases in elemental P content of periphyton cell tissues. Low concentrations of elemental P in periphyton cell tissues were highly correlated with the dominance of interior marsh filamentous blue-greens (Schizothrix calcicola and Scytonema hofmannii) and "alkaline" water diatoms (i.e., Mastogloia smithii v. lacustris, Cymbella rutnerii, Anomoeoneis vitrea and Synedra pahokeensis). The filamentous blue-green Microcoleus spp., the filamentous green Stigeoclonium sp., and the diatoms, Nitzschia amphibia, Navicula disputans, Navicula confervaceae, Gomphonema parvulum, and Nitzschia palea were highly correlated with marsh waters containing high concentrations of major ions, inorganic phosphorus, and nitrogen. Desmid relative abundance and periphyton species diversity indices were highest under acid, soft water conditions and high concentrations of elemental N in periphyton cell tissue (Table 6).

DISCUSSION

Major Ions, Algal Species Composition, and Diversity

Differences in the mineral content of WCA surface waters were closely related to the source of water for each marsh. Highest concentrations of major ions (bicarbonate, chloride, and calcium) occurred in WCA-2A which receives mineralized canal water inflows primarily from the three S-10 structures (See Figure 2). Due to the lack of interior perimeter canals, these waters are directly discharged across the northeast section of the marsh. Surface water inflows into WCA-2A represent a mixture of highly mineralized ground water and nutrient enriched canal water that drains the Everglades Agricultural Area south of Lake Okeechobee. Ground waters derived from this region are high in chloride and sodium, in contrast to calcium carbonate dominated ground water of WCA-3A and Everglades National Park (Flora and Rosendahl, 1981; Waller and Earle, 1975). Consequently, rainfall mixing with ground water derived from the Everglades Agricultural Area results

TABLE 6. CORRELATION COEFFICIENTS (R) FROM A STEPWISE MULTIPLE REGRESSION ANALYSIS OF DEPENDENT BIOLOGICAL VARIABLES VS. INDEPENDENT PHYSIO-CHEMICAL VARIABLES, WATER CONSERVATION AREA PERIPHYTON SURVEYS, 1978-1982

Dependent Variable (Biological)	Independent Variable (Physio-chemical)	R	% Variance Accounted	F Value
Periphyton elemental P content	Periphyton growth rates, water depth, marsh water PO_4	.83	73%	98.0
% <u>Schizothrix</u> - <u>Scytonema</u> rel. abund.	Periphyton N:P ratio, -periphyton elemental P content, -water depth	.806	65%	87.8
% Eutrophic diatom relative abundance: (Nitzschia amphibia, Navicula disputans, Navicula confervaceae, Gomphonema parvulum, Nitzschia palea)	Marsh water PO ₄ , periphyton elemental P content, inorganic N, -Magnesium, hardness	.728	54%	36.8
% Alkaline diatom relative abundance: (Mastogloia smithii v. lacustris Cymbella ruttneri, Anomoeoneis vitrea, Synedra pahokeensis, Cymbella minuta v. psuedogracilis)	Periphyton N:P ratio, -periphyton elemental P, -solar radiation	.714	51%	67.0
% <u>Microcoleus</u> relative abundance	Periphyton elemental P content, periphyton growth rates, marsh water inorganic nitrogen alkalinity	.690	48%	42.9
% Desmid relative abundance (Pleurotaenium, Cosmarium, Genicularia)	-Potassium, -magnesium, periphyton elemental N content	.513	26%	34.0
% <u>Stigeodonium</u> relative abundance	Marsh water PO ₄	.427	18%	42.7

in ratios of chloride:calcium that are relatively high (greater than 2.0) in WCA-2A (Table 1).

In contrast, surface waters in the interior of WCA-1 were low in dissolved minerals, poorly buffered, acid, and soft. The low mineral content of

these waters are a result of three factors: (1) WCA-1 is encircled by three levees (L-7, L-39, and L-40) which direct the majority of canal waters around the marsh perimeter. (2) Peat subsidence in the vicinity of these perimeter canals have created slightly lower ground level elevations around the peripheral edges of WCA-1

in comparison to ground levels in the interior marsh (Stephens and Johnson, 1951). The presence of perimeter canals, differences in ground level elevations, and the restriction of flow across the marsh to a large extent limits interior WCA-1 marsh water from mixing with mineralized canal water. (3) The high water holding capacity and capillary action of the deeper WCA-1 peat soils (up to 4 meters in depth) effectively holds large volumes of surface water (rainfall) above the main ground water mass. Together these factors effectively isolate interior WCA-1 marsh water from mineralized canal and ground water influences with the majority of its surface water being obtained directly from rainfall. Since rainfall contains relatively low concentrations of dissolved minerals, surface waters in the interior of WCA-1 are also low in electrolyte concentrations.

Surface water alkalinity and hardness in the interior of WCA-3A was roughly one half that of WCA-2A. Water in this region of the Everglades has been described as the "calcium carbonate type" (calcium is the dominant anion) and is influenced primarily by rainfall and dissolution and weathering of the Biscayne Aquifer (Gleason, 1972; Gleason and Spackman, 1974; Flora and Rosendall, 1981). Typically these waters have low ratios (less than 1.0) of chloride:calcium (Table 1).

Seasonal variations in the mineral content of interior marsh surface waters were weakly correlated with marsh hydrological variables such as water depth, concentration of ions by evapotranspiration, dilution by rainfall, or hydroperiod length. The ionic character and volume of each marsh's source water appear to be the most important factors influencing seasonal major ion concentrations in the WCA.

Results of this study and those of Gleason and Spackman (1974) and Swift (1981) indicate that major ion concentration is the predominant factor controlling periphyton species composition and species diversity within wet prairie communities of the interior marsh. These results contrast with Browder's (1981) work in Everglades National Park where percent soil organic matter and hydroperiod were reported as the major factors controlling the taxonomic composition of Everglades periphyton. Marsh waters of low ionic content (interior of WCA-1) favored development of an abundant desmid and filamentous green algae (Chlorophyceae) flora. These ion poor waters had a rich desmid flora, with over 120 species reported. As a result, low ionic content marsh waters were correlated with high species diversity. These results parallel earlier work in the WCA's (Swift, 1981) and compare well with previous studies of desmid ecology. Prescott (1952), Ruttner (1953), Hutchinson (1967), Moss (1972), Whitford and Schmaucher (1973), Wetzel (1975) and Woeklering (1976) report that desmids reach maximum

development in waters of low ionic content; especially in acid pH, low calcium, and magnesium habitats. Ruttner (1953) suggests that the electrolytic content (especially calcium carbonate) of the water appears to be associated with cell membrane permeability. Under hardwater conditions, desmid cells may become flooded with salts that they are unable to remove. As a result, desmids are less commonly found in alkaline, hard water habitats and are generally restricted to acid, low ionic content waters.

In contrast, interior marsh sites in WCA-2A, WCA-3A and the peripheral marsh of WCA-1 supported a filamentous blue-green flora dominated by Schizothrix calcicola, Scytonema hofmannii and an assemblage of diatoms indicative of hardwater environments (i.e., Mastogloia smithii v. lacustris). Filamentous blue-greens were the dominant algal group colonizing glass slides in waters containing moderate to high concentrations of major ions (bicarbonate, chloride, and calcium) and low nutrient content. These algae closely resembled the "calcareous periphyton" described by Van Meter (1965, 1973) and Gleason and Spackman (1974) for Everglades National Park. Calcareous periphyton refer to bluegreen algae communities which coat submerged macrophyte vegetation with algal filaments that are encrusted with calcium carbonate crystals. Floating mats of calcareous filamentous blue-greens attached to Utricularia (bladderwort) plants were a common feature of the WCA-2A and WCA-3A marshes during late summer and fall. In wet prairie communities, blue-green algae formed wooly, felt-like coatings or "cylinders" on submerged plant surfaces. During dry periods these algal communities desiccate and return organic matter and precipitated CaCO3 to the marsh sediment.

Increased sampling frequencies during 1980-82 revealed several trends that were not evident in earlier work (Swift, 1981). Filamentous blue-greens (Schizothrix and Scytonema) showed distinct seasonal patterns of growth and species succession. Maximum periphyton growth and abundance occurred during September-October of each year, near the end of the Minimum growth and abundance wet season. occurred during January-March. In earlier work, diatoms were not considered important members of WCA periphyton due to their relatively low biomass contribution to the total community. Results of this study indicate that diatoms play a much more important role at mineralized interior marsh sites, and frequently represent one third to one half of the total community by volume. Diatoms (e.g., Anomoeonis vitrea) were important pioneer algal species at several interior sites and rapidly recolonized submersed substrates following reflooding of the marsh after an extended dry period. Four diatom species recorded in this study represent new species. The fact that these species were present as abundant or common components of the WCA periphyton community demonstrates the lack of research that has been conducted regarding algal systematics in the Everglades region. In addition, two filamentous green algae (Mougetia and Spirogyra) were much more widely distributed throughout the WCA marsh than had been previously reported, and were seasonally important at both hard and soft water interior marsh sites.

Nutrients, Algal Growth Rates, and Nutritional Requirements

Results of this study support earlier observations concerning the nutrient status of the three WCA marshes: (a) during periods of inundation, inorganic forms of N and P at interior marsh sites remain low; (b) rainfall represents a major source of nutrients for interior marsh sites; (c) periphyton growth rates and elemental P concentrations within algal cell tissues are low at interior marsh sites suggesting that phosphorus represents the growth limiting nutrient within the interior marsh; (d) highest nutrient concentrations were recorded from peripheral marsh areas that are enriched by canal water inflows (sites B-2 and B-5); (e) algal communities exposed to nutrient enriched waters developed a "specialized" community of pollution tolerant species (i.e. Microcoleus spp. Oedogonium spp.) that have high growth rates, and high concentrations of elemental P within cell tissues.

Seasonal monitoring of interior marsh sites showed that hydrological events play an important role in recycling nutrients in the Water Conservation Areas. Inorganic nitrogen concentrations increase dramatically in the late dry season as water levels decline and approach the litter zone. These increases are the result of organic matter decomposition and release of ammonia and nitrates into the shallow water. Large releases of ammonia and nitrate are a common phenomenon in marshes which experience periodic exposure of soils that are normally inundated (Klopatek, 1978; Richardson et. al., 1978; Worth, 1983). Changes in water levels may result from natural climatic fluctuations or from man-induced changes, such as a marsh drawdown to re-establish the growth of certain aquatic plants. Release of phosphorus from interior marsh sediments into the water column was low in comparison to previous investigations of marsh reflooding (Kadlec, 1978; Klopatek, 1978; Federico et. al., 1978). Although these results are not consistent with previous studies of marsh reflooding, they compare well with Worth's (1983) data (ortho-PO₄ concentrations ranging from 2-4 µg/l) collected at interior WCA-2A sites during the same time period. Interior WCA marsh sites showed no major "first flush" effects (i.e., release of phosphorus from soils after reflooding) as has been described for Chandler Slough, Florida (Federico

et. al., 1978) and several other temperate marsh ecosystems (Richardson et. al., 1978).

Seasonal monitoring of periphyton growth rates at both interior and peripheral marsh sites showed contrasting results. High periphyton growth at nutrient enriched peripheral marsh sites were highly correlated with increases in the phosphorus content of marsh surface waters. Review of recent data collected from the northeast section of WCA-2A at Site B-5 clearly shows that S-10 discharges presently affect a much larger area of the marsh than previously reported in a preliminary 1978-79 survey (Swift, 1981). During 1978-1980, total dissolved P levels in water at B-5 were very low, averaging about 7 µg P/l. Algae growth rates were also low and contained low concentrations of elemental P within their cell tissues. During the drought of 1981, fire played an indirect but important role in modifying marsh water nutrient chemistry at Site B-5. Controlled burns of marsh vegetation in the northeast section of WCA-2A were conducted by the Florida Game and Fish Commission in the summer of 1981. This temporary reduction in sawgrass density permitted increased water flow across the marsh. Increased sheet flow, coupled with heavy autumn rains and high discharges through the S-10 structures allowed nutrient enriched canal water to penetrate the marsh much deeper than previously recorded. Biological response to these nutrient increases included (a) major shifts in periphyton species composition favoring development of algal communities tolerant of nutrient enriched conditions; (b) biostimulation (increased growth rates) of algal populations; and (c) increases in algal elemental P content.

Seasonal growth rates at interior marsh sites followed the classic "J" shape growth curve. Growth curves of this type are typical of algae populations which undergo rapid growth on a seasonal basis (i.e., "seasonal bloom" conditions). Kormondy (1969) indicates that the J shaped growth curve is indicative of unrestricted, rapid, exponential growth up to the limits of the environment. As these limits are exhausted (nutrients) or imposed (reduced light, temperature, or drought), the population declines to a reduced equilibrium level. Interior WCA periphyton communities exhibited low but well defined seasonal population maximums and minimums with chlorophyll a concentrations rarely exceeding 2.5 mg chl a · m-2 · wk-1. This suggests these microorganisms exist within relatively well-defined (low) limits of nutrient concentration. Seasonal fluctuations in algal growth at interior marsh sites were only weakly correlated with surface water nutrient concentrations. Seasonal growth rates were more related to variations in temperature, light, and water levels.

Periphyton elemental P concentrations at interior marsh sites were very low in comparison to

reported "critical concentrations" required for normal aquatic macrophyte and algae growth (Table 7). Supplies of phosphorus in periphyton tissue were comparable to levels reported from <u>Utricularia</u>-periphyton communities in the Okefenokee Swamp (Bosserman, 1979) and from Everglades aquatic plants and organic peat soils (Davis and Harris, 1978; Stewart and Ornes, 1975; Volk et. al., 1975). Low concentrations of elemental P in periphyton cell tissue, low periphyton growth rates, and the low phosphorus content of marsh surface waters and soils suggest that phosphorus is the nutrient limiting element at interior WCA-2A and WCA-3A sites.

Elemental P content of periphyton communities collected from ion poor waters (interior of WCA-1) were 2-3 times higher than those recorded from calcium rich interior marsh sites. Since rainfall probably represents the major source of P to interior marsh sites, the question may be asked, "Why do acid, soft water periphyton communities contain higher levels of P when compared to algae collected from calcium rich environments?" One explanation suggested is that under conditions of high calcium carbonate precipitation, "coprecipitation" of phosphorus as hydroxylapitite Ca₂(OH)(PO₄)³ may occur naturally removing available P from the water column (Wetzel, 1975). Consequently, the calcium content of marsh surface waters may have a significant effect on amounts of P available for plant growth at calcium rich interior marsh sites. In temperate marl lakes calcium carbonate precipitation is a significant factor in the regulation of algal productivity both by thispagereserved for Table 7 adsorption of liable organic substrates and through interaction with phosphate (Otsuki and Wetzel, 1972). Coprecipitation of P may partially explain the observed low P content of marsh surface waters and soils, low algal growth rates and low elemental P content of periphyton observed throughout this study at calcium rich interior marsh sites.

Increases in elemental P content of periphyton at peripheral marsh sites were highly correlated with increases in nutrient content of marsh surface waters and periphyton growth rates. Periphyton elemental P content was well above the 0.2 percent "critical concentration" level reported necessary for producing maximum growth responses in blue-green algae (Table 7). The high phosphorus content of peripheral marsh periphyton indicates these algae are storing phosphorus in excess of their growth requirements (i.e., luxury consumption) in response to S-10 nutrient enrichment.

Concentrations of nitrogen in periphyton tissue were comparable to ranges reported for <u>Utricularia</u>-periphyton mats from the Okefenokee Swamp, and ranges reported for most aquatic macrophytes. The nitrogen content of WCA periphyton were always

above Vallentyne's (1974) 0.7 percent N "lower limit" at which growth may occur in freshwater plants and algae (Table 7). This suggests that nitrogen was present in adequate supply to permit algal growth. However, nitrogen content of the majority of samples collected was usually below Gerloff and Skoog's 4.0 percent "critical concentration" necessary to produce a maximum growth response in laboratory cultures of blue-green algae. Although Gerloff and Skoog demonstrated nitrogen storage in blue-greens, our field data showed periphyton nitrogen levels to be weakly correlated with marsh water nutrient chemistry. Two incidences of nitrogen uptake and storage were reported during low water conditions at Stations A-3 and the 3-4 gauge during 1981. Filamentous green algae (Mougeotia sp. and Spirogyra sp.) communities showed rapid, short term increases in their biomass and elemental N content in response to increases in inorganic nitrogen concentrations (primarily ammonia) during low water periods.

Indicator Organisms

Results of this study and review of the literature show that specific assemblages of periphyton can be used as fairly reliable indicators of water quality type within the Water Conservation Areas.

Periphyton communities dominated by the blue-greens Schizothrix calcicola and Scytonema hofmannii in association with the diatoms, Mastogloia smithii v. lacustris, Anomoeoneis vitrea, Cymbella rutnerii, Synedra pahokeenis sp. nov. and Cymbella minuta v. psuedogracilis represent low nutrient, hard water (calcium carbonate dominated) water quality conditions. Conversely, algal communities dominated by filamentous greens (Mougeotia spp.) and a rich desmid flora in close association with several species of diatoms (Cymbella amphioxys, Frustulia rhomboides v. saxonica, Anomoeoneis serians v. brachysira, Anomoeoneis serians, Eunotia neageli, and Stenopterobia intermedia) represent low pH (acid), soft water marsh conditions.

Algal communities dominated by large populations of the blue-green Microcoleus lyngbyaceus or the green alga, Oedogonium spp. in close association with the diatoms, Nitzschia amphibia, Nitzschia palea, Navicula disputans, Navicula confervaceae or Gomphonema parvulum provided good evidence that these waters have recently contained relatively high concentrations of inorganic phosphorus and nitrogen (Van Landingham, 1981; Reimer, 1962; Lowe, 1974).

Comparison of Peatland Marsh Water Chemistry

Water chemistry characteristics of peat forming marshes fall within two extremes: (a) ombrotophic peatlands (bogs) which receive their water supplies directly from precipitation, and (b) minerotrophic

	Elemental Nut	rient Concentration	-
tudy %N	%P	N:P Ratio	
Critical concentrations"			
roducing maximum growth			
esponse in <u>Microcystis</u> spp.			
b culture (Gerloff and			
koog, 1954).	4.0	0.2	20:1
ssential concentrations of			
I and P required for growth			
n freshwater plants			
Vallentyne, 1974).	0.7	0.08	7:1*
ranentyne, 1974).	0.7	0.00	7.1
langes of nutrient			
oncentrations for aquatic			
nacrophytes (Boyd, 1978).	1.46-3.95	.08-0.63	ND
Ontario, Canada			
Average periphyton nutrient			
ontent			
oligiotrophic lake	2.59	.06	43:1
fertilized lake	2.56	.18	14:1
Stockner and Armstrong, 1971)			
eriphyton growth in			
rtificial streams, averages	3.29-1.63	.2111	16:1
and standard deviations	3.2303		
Kevern and Ball, 1965)			
Okefenoke Swamp, Georgia			
langes of N and P content for			
<u>Itricularia</u> periphyton mats			
Bosserman, 1979)	1.73-3.09	.045075	38-41:1
Nater Conservation Areas, Fl.			
lutrient content of natural			
ubstrate periphyton (ranges)	0.00.0 = 0	040 004	20.74.46
Interior WCA-1 (Site A-3)	2.32-3.79	.048094	29-71:1(avg. 51:1)
Interior WCA-2A (217 gauge)	.94-3.57	.01042	60-205:1 (avg. 101:1)
Interior WCA-3A (3-4 and 3-7			
gauges)	1.32-3.22	.01041	59-183:1 (avg. 105:1)
Nutrient enriched Site B-2	2.21-3.60	.224419	6.5-13.4:1 (avg.8.6:1)
ND = No data			

peatlands (fens) which derive the majority of their waters from ground water sources. Most peat forming marshes can be placed somewhere within these two extremes since they receive various amounts of water from both rainfall and ground water.

approximately 40 carbon: 7 nitrogen: 1 phosphorus per 100 dry weight.

This classification scheme is useful to this study because the origin of the marsh's water directly affects its chemical properties and ultimately influences periphyton species composition within the marsh. Minerotrophic fens develop in marsh systems which drain alkaline soils or overlie mineral soils. Minerotrophic fens can also develop in marshes which are influenced by mobile ground water sources (either continuous or intermittant). In either case, minerotrophic fens are characterized as alkaline, hard water environments dominated by bicarbonate and calcium ions with circumneutral to basic pH, high specific conductivity and generally low concentrations of inorganic forms of nitrogen and phosphorus (Moore and Bellamy, 1974).

Water quality within interior marshes of Water Conservation Areas 2A, 3A, and the peripheral marsh of WCA-1 can be best described as minerotrophic. The dominant major ions of these marshes are bicarbonate, calcium, chloride, and sodium. Hydrogen ion concentrations (pH) are usually within the neutral to basic range, while average specific conductivity ranges from 333-1050 µmhos/cm. Inorganic forms of N and P were generally at or near detection levels throughout the year (with the exception of the onset of the dry season). Calcium carbonate precipitating species of blue-green algae (Schizothrix calcicola and Scytonema hofmannii) and the diatom complex comprised of Mastogloia smithii v. lacustris, Cymbella ruttneri, Synedra pahokeensis sp. nov. and Anomoeneis vitrea represented the characteristic microflora of WCA minerotrophic peat forming marshes.

Ombrotrophic peatlands (bogs) typically contain an interior raised bog (interior of marsh has a higher ground level than the peripheral marsh due to peat accumulation) and an accompanying perched water table. The accumulation of peat acts as a reservoir holding large volumes of water above the ground water mass. As a result, ombrotrophic peatlands are isolated from ground water inflows and derive the majority of their water from direct rainfall. Since rainfall is ion-poor, surface water of these marshes is also ion-poor and typically contain calcium concentrations of less than 5 mg/l and specific conductance less than 15 µmhos/cm (Moore and Bellamy, 1974). Ombrotrophic peatlands are acidic with pH values ranging from 3-4.5 as a result of the cation exchange properties of various Sphagnum species of mosses which dominate ombrotrophic situations (Cylmo, 1963). Desmids represent the characteristic microflora of ombrotrophic bog communities (Wetzel, 1975).

Water quality in the interior of WCA-1 represents an intermediate or transitional stage of marsh succession from minerotrophic fen to ombrotrophic bog. Marsh waters of interior WCA-1 are ion poor and contain approximately 2-3 times the concentration of electrolytes present in local rainfall. Surface waters are typically within the acid range (pH 5.7) with specific conductance averaging about 100

µmhos/cm, and calcium concentrations averaging less than 8 mg/l. Desmids, filamentous green algae, and certain diatom species indicative of acid, soft water conditions represent the dominant microflora of interior WCA-1.

Table 8presents a comparison of the general shift in ionic components of marsh waters in the transition from minerotrophic fen to ombrotrophic bog (after Wetzel, 1975) with comparisons of averaged data collected from interior sites from each of the three Water Conservation Areas.

European minerotrophic fens compared well with WCA-2A and WCA-3A with respect to pH, calcium, bicarbonate, potassium and magnesium concetrations. However, chloride and sodium concentrations in WCA-2A exceeded the European fen average approximately tenfold. Sulfate concentrations in the WCA's were lower than the European fen average.

Water quality data from WCA-1 compared well with European transitional fen averages but had significantly higher concentrations of dissolved minerals when compared to classic ombrotrophic raised bog environments.

Effects of Water Management on Everglades Periphyton

The predominant water management practice affecting the Water Conservation Areas today is the impoundment of surface water runoff derived from the Everglades Agricultural Area for flood control and water storage purposes. Future water management plans for south Florida may call for backpumping urban stormwater runoff, presently discharged eastward to tidewater, into the Water Conservation Areas (SFWMD, 1977, 1983). Benefits of these backpumping plans include increased water storage capacity for recharge of regional ground water supplies and protection of east coast well fields against salt water intrusion. Environmental concerns of these plans focus on the impact of increasing nutrient concentrations, water levels, and hydroperiod lengths within the Water Conservation Areas. One goal of this study was to address these impacts as they affect periphyton communities in the Water Conservation Areas.

Introduction of Nutrients as a Result of Backpumping.

Results of this study indicate that increases in marsh water N and P supplies (1) promote major changes in periphyton species composition, (2) reduce algal species diversity, (3) stimulate the growth of pollution tolerant algae, and (4) increase the P content of algal cells. Changes in periphyton species composition and reduced algal species diversity as a result of nutrient

Direction of Peatland Water Chemistry Succession Minerotrophic fen (surface waters influenced by ground water inflows) Transitional fen (interior marsh receives direct rainfall, continued peat accumulation diverts ground water inflows accumulation diverts of basin) Ombrotrophic "raized" Bog (perched water table isolated from ground water) ND = No Data Available	A COMPARISON OF AVERAGE IONIC COMPOSITION OF WESTERN EUROPEAN/SCANDANAVIAN P SURFACE WATERS WITH AVERAGED DATA FROM THE THREE WATER CONSERVATION AREAS	COMPOSITION OF WESTERN EUROPEAN/SCANDANAVIAN PEATLAND D DATA FROM THE THREE WATER CONSERVATION AREAS	N EUROPEAN/S WATER CONSE	CANDANA	VIAN PEA	TLAND		
Western European(a) mineroptrophic fens WCA-2A WCA-3B Transitional fen(b) S(Scandanavia) Transitional mire(a) S(Western Europe, Type No. 4) WCA-1 Substant Europe, Type No. 4) WCA-1 Substant Europe, Type No. 4) WCA-1 Substant Europe, Type No. 4) Substant Europe, Type No. 4) WCA-1 Substant Europe, Type No. 4) Substant Europe, Type No. 4)	Hď	Conduc- tivity Ca++ (<u>lhos/cm) (mg/l)</u>	+ HCO ₃ = (1)	Na + (mg/!)	Cl [.] (mg/l)	Mg + + (mg/l)	K + (mg/l)	\$04 - (mg/l)
WCA-2A WCA-3B Transitional fen(b) Scandanavia) Transitional mire(a) Swestern Europe, Type No. 4) WCA-1 Substance (1)	7.6	ND 80	195	11.5	17.7	10.9	8 .	48
WCA-3B WCA-3A Transitional fen(b) 5 (Scandanavia) Transitional mire(a) 5 (Western Europe, Type No. 4) WCA-1 Subprotrophic(1) 3 Bog (<u>Sphagnum</u> dominated)	7.1	1050 67	245	121	174	28	6.5	39
WCA-3A 7 Transitional fen(b) 5 (Scandanavia) Transitional mire(a) 5 (Western Europe, Type No. 4) WCA-1 5 Ombrotrophic(1) 3. Bog (<u>Sphagnum</u> dominated)	7.4	566 54	184	57	64	6	2.4	9
Transitional fen(b) 5 (Scandanavia) Transitional mire(a) 5 (Western Europe, Type No. 4) WCA-1 5. WCA-1 5. Ombrotrophic(1) 3. Bog (<u>Sphagnum</u> dominated)	7.0	333 47	132	18	28	4	1.1	12
Transitional mire(a) 5 (Western Europe, Type No. 4) WCA-1 5 Ombrotrophic(1) 3 Bog (<u>Sphagnum</u> dominated)	en(b) 5.8	ND 18	45	7	3.5	0.2	6.0	4.1
WCA-1 Ombrotrophic ⁽¹⁾ Bog (<u>Sphagnum</u> dominated)	a) 2	D 14	20	11.5	17	2.4	9.1	24
Ombrotrophic(1) 3 Bog (<u>Sphagnum</u> dominated)	5.7	112 6	16	12	22	1.5	0.6	6
ND = No Data Available		<15 0.8	-	5	1.4	9.0	4	4.8
:								
(1) Moore and Bellamy, 1974								
(2) Sjors, 1950								

enrichment may have important long term effects on marsh fish and invertebrate fauna. Inasmuch as periphyton represent a significant component of the marsh food web, major shifts in species composition at the primary producer level may ultimately influence both the species composition and biomass of higher trophic level organisms. Changes in food item size, reduction in food selection choices, or changes in food nutritive value denote the possible effects of nutrient enrichment on herbivorous invertebrates and fish fauna.

Aquatic ecosystems containing high species diversity generally contain more complex pathways of energy flow between trophic levels (i.e, have more complex food webs or food chains) and represent more stable ecosystems in comparison to simpler, less diverse communities (Odum, 1971). Reduction of periphyton species diversity as a result of nutrient enrichment may ultimately reduce ecosystem stability by reducing the number of food item choices available to grazing invertebrates.

Increasing Marsh Water Ionic Content as a Result of Backpumping.

Results show that periphyton species composition at interior marsh sites are also strongly influenced by the ionic content of marsh surface waters. Discharge of mineralized canal water (high calcium carbonate content) across WCA-2A, WCA-3A, and the peripheral WCA-1 marsh will have little effect on resident bluegreen/diatom communities because these marshes are already hard water ecosystems. However, discharge of mineralized canal waters across the interior of WCA-1 would significantly change the periphyton flora and water quality characteristics of the marsh interior from an acid, soft water marsh dominated by desmids and filamentous greens to an alkaline, hard water ecosystem dominated by filamentous blue-greens.

In practice, waters that could potentially be backpumped into the WCA's would almost always contain high concentrations of N and P as well as calcium carbonate. It is the author's opinion that excess N and P concentrations have had far greater impacts on the Everglades periphyton community, as a whole, in comparison to increases in the calcium carbonate content of WCA surface waters.

LITERATURE CITED

Allen, R.J. Jr. 1978-1982. Climatological Report. University of Florida, IFAS, Agricultural Research and Education Center, Belle Glade, Florida.

American Public Health Association (APHA). 1975. Standard Methods for the Examination of Water and Waste Water, 14th Ed., Washington, D.C.

Arnold, D.E. 1971. Ingestion, Assimiltion, Survival, and Reproduction by <u>Daphnia pulex</u> Fed Seven Species of Blue-green Algae. Limnol. Oceanogr. 16:906-920.

Black, C.A. (ed.). 1965. Methods of Soil Analysis, Agron. No. 9. Amer. Soc. of Agron. Madison, Wisc. p. 770.

Bosserman, R.W. 1979. The Hierarchical Integrity of <u>Utricularia</u>-periphyton Microecosystems. Ph.D. Dissertation, University of Georgia, Athens, Ga.

Boyd, C.E. 1978. Chemical Composition of Wetland Plants. In: D.F. Whigham, R.L. Simpson, and C.G. Jackson, eds. Freshwater Wetlands. Academic Press. New York, pp. 155-166.

Browder, J.A. 1981. Perspective on the Ecological Causes and Effects of the Variable Algal Composition of Southern Everglades Periphyton. South Florida Research Center, Report T-643, Everglades National Park, Homestead, Florida. p. 110.

Carter, M.R., L.A. Burns, T.R Cavinder, K.R. Dugger, P.L. Fore, D.B.Hicks, H.F. Revels, and T.M. Schmidt. 1973. Ecosystems Analysis of the Big Cypress Swamp and Estuaries: Environmental Protection Agency, Ecological Report No. DI-SFEP-74-51, p. 375.

Craighead, F.C., Sr. 1971. The Trees of South Florida. Vol. 1. The Natural Environments and their Succession, University of Miami Press, Coral Gables, p. 66.

Cylmo, R.S. 1963. Ion Exchange in <u>Sphagnum</u> and its Relation to Bog Ecology. Ann. Bot. N.S. 27:309.

Davis, J.H. Jr. 1943. The Natural Features of Southern Florida, Especially the Vegetation and the Everglades. Geol. Bull. No. 25, Fla. Dept. Conserv. Fla. Geol. Surv. Tallahassee. p. 311. 1946. The Peat Deposits of Florida, their Occurrence, Development, and Uses. Geol. Bull. No. 30. Fla. Dept. Conserv. Fla. Geol. Surv. Tallahassee. p. 247.

Davis, S.M. 1981. Mineral flux in the Boney Marsh, Kissimmee River. 1. Mineral retention in relation to overland flow during the three year period following reflooding. Technical Publication 81-1, Resource Planning Dept. South Florida Water Management District, West Palm Beach, FL.

Davis, S.M. and L.A. Harris. 1978. Marsh Plant Production and Phosphorus Flux in Everglades Conservation Area 2: In: Environmental Quality through Wetlands Utilization. M.A. Drew (ed.). Symposium Proc. Restoration of the Kissimmee River and Taylor Creek - Nubbin Slough Basin, February 28 - March 2, 1978. Tallahassee, Fla.:171-181.

Drouet, F. 1963. Ecophenes of Schizothrix calcicola. Proc. Acad. Nat. Sci., Philadelphia. 15(9):261-281.

Oscillatoriaceae. Acad. Nat. Sci. Philadelphia, Monog. No. 15.

1973. Revision of the Nostocaceae with Cylindrical Trichomes. Hafner Press, New York.

Drouet, F. and W.A. Daily. 1965. Revision of the Coccoid Myxophyceae. Butler Univ. Botany Studies 12:218.

Evans, D. and J.G. Stockner. 1972. Attached Algae on Artificial and Natural Substrates in Lake Winnipeg, Manitoba. J. Fish Res. Board Can. 29:31-44.

Environmental Protection Agency. 1976. A Manual of Methods for Chemical Analysis of Water and Wastes. Office of Technology Transfer, Washington, D.C.

Federico, A.C., J.F. Milleson, P.S Millar, and M. Rosen. 1978. Environmental Studies in the Chandler Slough Watershed. Tech. Pub. #78-2, South Florida Water Management District, West Palm Beach, FL.

Flora, M.D. and P.C. Rosendahl. 1981. Specific Conductance and Ionic Characteristics of the Shark River Slough, Everglades National Park, Florida. Report T-165. Everglades National Park, Homestead, Florida. p. 55.

Gerloff, G.C. and F. Skoog. 1954. Cell Contents of Nitrogen and Phosphorus as a Measure of their Availability for Growth of <u>Microcystis aeruginosa</u>. Ecology 35:348-353.

Gleason, P.J. 1972. The Origin, Sedimentation and Stratigraphy of a Calcite Mud Located in the Southern Freshwater Everglades. Ph.D. Thesis, College of Earth and Mineral Sciences, Pennsylvania State University, Penn. p. 355.

Gleason, P.J. 1974. Chemical Quality of Water in Conservation Area 2A and Associated Canals. Technical Publication 74-1, February 1974. Resource Planning Department, South Florida Water Management District, West Palm Beach, FL.

Gleason, P.J., A.D. Cohen, W.G. Smith, H.K. Brooks, P.A. Stone, R.L. Goodrick, and W. Spackman, Jr. 1974. The Environmental Significance of Holocene Sediments from the Everglades and Saline Tidal Plain. In: P.J. Gleason (ed.), Environments of South Florida: Present and Past, pp. 287-341, Memoir 2, Miami Geological Society, Miami, FL.

Gleason, P.J. and W. Spackman, Jr. 1974. Calcareous Periphyton and Water Chemistry in the Everglades. In: P.J. Gleason (ed.), Environments of South Florida: Present and Past, pp. 146-181, Memoir 2, Miami Geological Society, Miami, FL.

Goldstein, J., M. Dowling, and R. Benner, (In press). 1980. Nitrogen Fixation in the Florida Everglades. Nat. Science Found. Grant, Biology Dept., Antoich College, Yellow Springs, Ohio.

Hunt, B.P. 1961. A Preliminary Survey of the Physico-chemical Characteristics of Taylor Slough with Estimates of Primary Productivity. Report to Supt. of Everglades National Park, Jan. 12, 1961, pp. 1-26 (Mineo.).

Hustedt, F. 1930. Bacillariophyta (Diatomeae). In: A. Pascher (ed.) Die Susswasser-Flora Mitteleuropas, Bd. 10. Gustav Fischer, Dena, pp. 1-466.

Hutchinson, G.E. 1957. A Treatise on Limnology. Vol. 1, Geography, Physics and Chemistry. John Wiley and Sons, New York, p.1015.

n Limnology. Vol. 2, Introduction to Lake Biology and the Limnoplankton. John Wiley and Sons, New York, p. 1115.

. 1973. Eutrophication. American Sci. Vol. 61, pp. 269-279.

Kadlec, R.H. and J.A. Kadlec. 1978. Wetland Functions and Values: The State of our Understanding. American Water Resources Association, November 1978, pp. 436-456.

Kim, Y.C. 1967. The Desmidiaceae and Mesotaeniaceae in North Carolina. Ph.D. Thesis, Botany Dept., North Carolina State University, Raleigh, N.C. (University Microfilms, Inc., Ann Arbor, Mich.).

King, D.L. and R.C. Ball. 1966. A Qualitative and Quantitative Measure of Aufwuchs Production, Tr. Amer. Microscop. Soc. 82:232.

Klopatek, J.M. 1978. Nutrient Dynamics of Freshwater Riverine Marshes and the Role of Emergent Macrophytes. In: R.E. Good, D.F. Whigham, and R.L. Simpson (eds.). Freshwater Wetlands, Ecological Processes and Management Potential. Academic Press, New York, pp. 195-216.

Kolopinski, M.C. and A.L. Higer. 1969. Effects of Mineralized Artesian Water on the Freshwater Biota of Taylor Slough, Everglades National Park. U.S. Geological Survey Open File Report, July 1969.

Kormondy, E.J. 1969. Concepts of Ecology. Prentice-Hall, Inc., Englewood Cliffs, New Jersey. p. 209.

Lowe, R.L. 1974. Environmental Requirements and Pollution Tolerance of Freshwater Diatoms. U.S. Environmental Protection Agency - 670/4-74-005. p. 333.

Lutz, J. 1977. Water Quality Characteristics of Several Southeast Florida Canals. Technical Pub. #77-4. Resource Planning Dept., South Florida Water Management District, West Palm Beach, FL. p. 83.

McPherson, B.F., B.G. Waller, and H.C. Mattraw. 1976. Nitrogen and Phosphorus Uptake in the Everglades Conservation Areas, Fla., with Special Reference to the Effects of Backpumping Runoff. Water Resources Investigations 76-29. U.S. Geological Survey, Tallahassee, Florida.

Millar, P.S. 1980. Water Quality Analysis in the Water Conservation Areas. Technical Memorandum Interim Progress Report, May 1981. Resource Planning Dept., South Florida Water Management District, West Palm Beach, Florida.

Moore, P.D., and D.J.Bellamy. 1974. Peatlands. Springer-Verlag, New York Inc., New York. p. 221.

Moss, B. 1972. The Influence of Environmental Factors on the Distribution of Freshwater Algae: An Experimental Study. I. Introduction and the Influence of Calcium Concentrations. J. Ecology, 60:917-932.

Odum, E.P. 1971. Fundamentals of Ecology. W.B. Sanders Co., Philadelphia, p. 547.

Otsuki, A. and R.G. Wetzel. 1972. Coprecipitation of Phosphates with Carbonates in a Marl Lake. Limnol. Oceanogr., 17:763-767.

Palmer, M.C. 1969. A Composite Rating of Algae Tolerating Organic Pollution. Journal of Phycology. Vol. 5, No. 1, pp. 78-82

Patrick, R. 1977. Ecology of Freshwater Diatoms and Diatom Communities. In: Werner, D. (ed.). The Biology of Diatoms. University of California Press, Berkeley, Ca.

Patrick, R. and C.W. Reimer. 1966 and 1973. The Diatoms of the United States. Vol. 1. Academy of Natural Sciences, Philadelphia, Monograph No. 13, p. 688.

the United States. Vol. 2. Part 1. Academy of Natural Sciences, Philadelphia Monograph No. 13(2) p.213.

Prescott, G.W. 1952. Algae of the Western Great Lakes Area. W.C. Brown Co., Dubuque, Io. p. 977.

Reimer, C.W. 1962. Diatoms and their Physicochemical Environment. In: Fogg, G.E. 1962. Environmental Requirements of Plankton Algae and their Effects on Water Quality. Third Seminar, Biological Problems in Water Pollution.

Reynolds, C.S. and A.E. Walsby. 1975. Waterblooms. Biol. Rev. 50:437-481.

Richardson, C.J., D.L. T ilton, J.A. Kadlec, J.P.M. Chami, and W.A. Wentz.. 1978. Nutrient Dynamics of Northern Wetlands Ecosystems. In: Freshwater Wetlands: Ecological Processes and Management

Potential (R.E. Good, D.F. Whigman, and R.L. Simpson, eds.). Academic Press, pp. 217-241.

Ricklefs, R.E. 1979. Ecology. Chiron Press, 2nd Edition, New York and Concord. p. 966.

Round, F.E. 1965. The Biology of Algae. St. Martin's Press, New York. p. 269.

Ruttner, F. 1953. Fundamentals of Limnology, University of Toronto Press, Toronto, Canada.

Schomer, S.N. and R.D. Drew. 1982. An ecological characterization of the Lower Everglades, Florida Bay, and the Florida Keys. U.S. Fish and Wildlife Service, Office of Biological Services, Washington, D.C. FWS/OBS-82/58.1.

Shannon, C.E. and W. Weaver. 1963. The Mathematical Theory of Communication. University of Illinois Press, Urbana. p. 117.

Sjors, H. 1950. On the Relation Between Vegetation and Electrolytes in North Swedish Mire Waters. Oikos 2:241-258.

Smith, G.M. 1950. The Freshwater Algae of the United States, 2nd Edition. McGraw-Hill Book Co., New York. p. 719.

Smith, W.G. 1968. Sedementary environments and environmental change in the peat-forming area of South Florida. Ph.D. dissertation, College of Earth and Mineral Sciences, Pennsylvania State University.

Snedecor, G.W., and W.G. Cochran. 1967. Statistical Methods. Iowa State University Presss, Ames, Iowa, p. 593.

Stephens, J.C. 1956. Subsidence of Organic Soils in the Florida Everglades. Soil Sci. of Am. Proc. Vol. 20, pp. 77-80.

Stephens, J.C. and L. Johnson. 1951. Subsidence of Organic Soils in the Upper Everglades Region of Florida. Soil Sci. of Fla. Proc., Vol. XI, pp. 191-237, Gainesville, FL.

Stewart, K.K. and W.H. Ornes. 1975. The Autecology of Sawgrass in the Florida Everglades. Ecology 56:162-171.

Stockner, J.G. and F.A.J. Armstrong. 1971. Periphyton of the Experimental Lakes Areas (ELA), Northwestern Ontario. J. Fish Res. Bd. Canada 28: pp. 215-229.

Strickland, J.D.H. and T.R. Parsons. 1968. A Practical Handbook of Seawater Analysis. Bull. Fish. Res. Board Can. 167:p. 310.

Tabb, D.C., T.R. Alexander, T.M. Thomas, and N. Maynard. 1967. The Physical, Biological, and Geologial Character of the Area South of C-111 Canal in Extreme Southeastern Everglades National Park, Florida. Report to the U.S. National Park Service, Institute of Marine Sciences, University of Miami, pp. 25-37.

Swift, D.R. 1981. Preliminary Investigation of Periphyton and Water Quality Relationships in the Everglades Water Conservation Areas. Technical Publication 81-5, December 1981. Resource Planning Department, South Florida Water Management District, West Palm Beach, Florida. p. 83.

Vanlandingham, S.L. 1981. Guide to the Identification, Environmental Requirements and Pollution Tolerance of Freshwater Blue-green Algae (Cyanophyte). Report No. EPA-600/3-82-073. Environmental Protection Agency, Cincinnati, Ohio. p. 341.

Vallentyne, J.R. 1974. The Algal Bowl - Lakes and Man. Ottawa, Misc. Special Publ. 22, Dept. of the Environment, p. 185.

VanMeter, N. 1965. Some Quantitative and Qualitative Aspects of Periphyton in the Everglades. Unpublished M.S. Thesis, University of Miami, Coral Gables, Florida.

VanMeter-Kasonof, N. 1973. Ecology of the Macroalgae of the Florida Everglades. Part I - Environment and Some Aspects of Freshwater Periphyton, 1959-1963. Nova Hedwigia 24:619-664.

Volk, B.G., S.D. Schemnitz, J.F. Gamle, and J.B Sartain. 1975. Baseline Data on Everglades Soil-Plant Systems: Elemental Composition, Biomass and Soil Depth. In: Howell, F.G., Gentry, J.B. and Smith, M.H. (eds). Mineral Cycling in Southeastern Ecosystems. Available as CONF-740513, NTIS. U.S. Dept. of Commerce, Springfield, Va.

Waller, B.G. and J.E. Earle. 1975. Chemical and Biological Quality of Water in Part of the Everglades, Southeastern Florida. U.S. Geological Survey Water Resources Investigations, Tallahassee, Florida.

Wetzel, R.G. 1975. Limnology. W.B. Saunders Company, Philadelphia, Pa. p. 743.

Whitford, L.A. and G.J. Schumacher. 1973. A Manual of the Freshwater Algaue in North Carolina. Agri. Exper. Station, Raleigh, N.C.

Wilson, S.U. 1974. Metabolism and Biology of a Blue-green Algal Mat. Unpublished M.S. Thesis, University of Miami, Miami, Florida.

Woelkering, W.J. 1976. Wisconsin Desmids. I. Aufwuchs and Plankton Community of Selected Acid Bogs, Alkaline Bogs, and Closed Bogs. Hydrobiologia 48(3): pp. 209-232.

Wood, E.J.F. and N.G.Maynard. 1974. Ecology of Micro-algae of the Florida Everglades. In: P.J. Gleason (ed.), Environments of South Florida: Present and Past. pp. 123-145.

Worth, D. 1983. Preliminary Environmental Responses to Marsh Dewatering and Reduction in Water Regulation Schedule in Water Conservation Area 2A. Technical Publication 83-6, September 1983. South Florida Water Management District, West Palm Beach, Florida. APPENDIX A SPECIES LIST

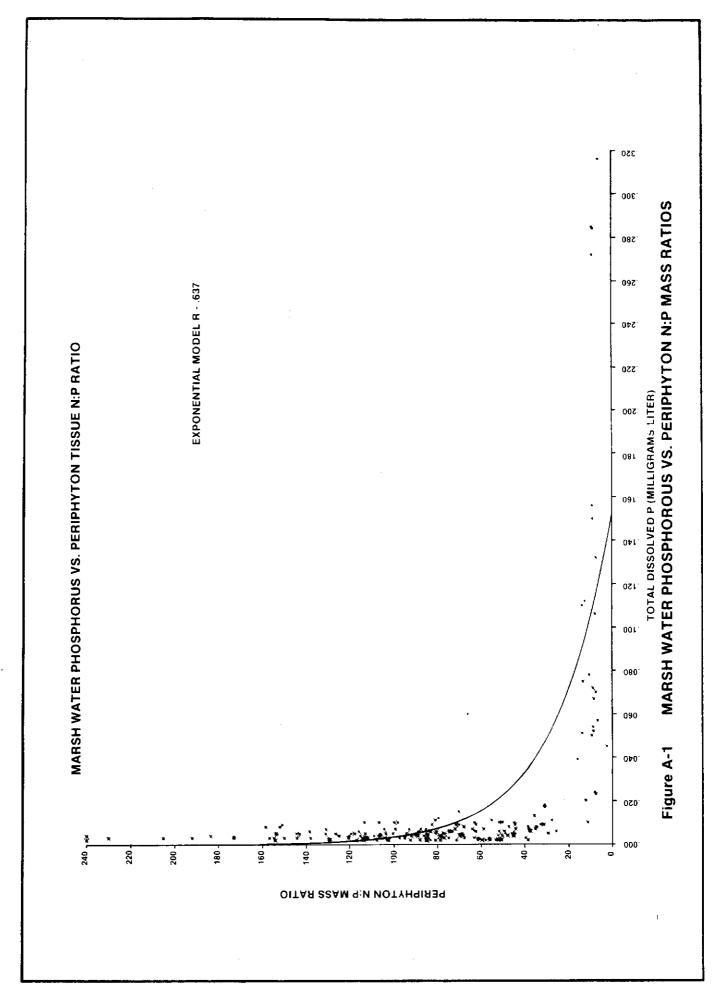
SYSTEMATIC LIST OF PERIPHYTON SPECIES

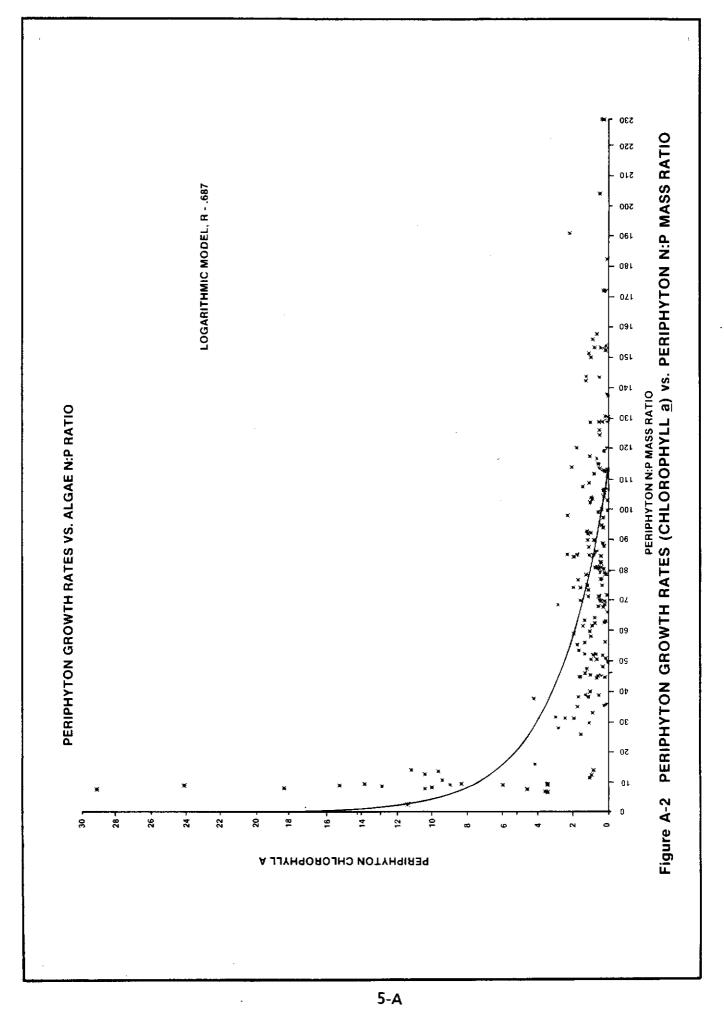
Division:			
	Myxophyceae (Blue-Greens) ^a	Families: Mesotaeniaceae	Spondylosium planum
	Family: Chroococcaceae	and Desmidiaceae (con't.)	Staurastrum alternans
	Agmenellum quadruplicatum	Cosmarium alpestre	Staurastrum americanum
	Anacystis dimidiata	Cosmarium angulosum	Staurastrum arachne
	Anacystis montana	v. concinnum	Staurastrum cf. comptum
	Coccochloris elabens	Cosmarium bisphaericum	v. pinnatiforme
	Gomphospheria spp.	Cosmarium blyttii	Staurastrum crytocerum
	Johannesbaptistia pellucida	Cosmarium circulare	Staurastrum dejectum
	Family: Oscillatoriaceae	Cosmarium comminsurale	Staurastrum dickiei
		V. Crassum	v. rhomboideum
	Microcoleus lynbyaceus		Staurastrum forficulatum
	Oscillatoria lutea	Cosmarium of contractum	v. eximium
	Oscillatoria princeps	Cosmarium of costatum	Staurastrum lativenter
	Schizothrix calcicola	Cosmarium elegantissimum	
	Spirulina princeps	v. simplicus	Staurastrum leptocladium
	Spirulina subsalsa	Cosmarium excavatum	Staurastrum inversenii
	Family: Nostocaceae	f. duplomajor	Staurastrum johnsonii
	Anabaena spp.	Cosmarium exiguum	Staurastrum ophiura
	Scytonema hofmannii	Cosmarium of galeritum	v. cambricum
	Family: Stigonemataceae	Cosmarium cf. granatum	Staurastrum paradoxum
	Stigonema panniforme	Cosmarium hammeri	v. harvulli
B	and the same	Cosmarium cf. impressulum	
: מסופועוט	Chlorophyceae (Greens) ^b	Cosmarium margaritatum	Staurastrum triundulatum
uraer	: Volvocales	Cosmarium of, moniliforme	Tetmemorus granulatus
	Carteria spp.	Cosmarium notable	Triploceras gracile
	Chlamydomonas spp.	Cosmarium orthostichum	Triploceras verticillatum
. .	_Eudorina elegans	Cosmarium cf. punctulatum	
Order	: Tetrasporales	Cosmarium pyramidatum	v. fissum
	Elakatothrix gelatinosa	Cosmarium pyramidatum	Xanthidium cristatum
_	Gleocystis gigas	v. convexum	v. enicinatum
Order		Cosmarium of.	Xanthidium f. micronatum
	Chaetopleltis obicularis	schliephackeanum	Xanthidium sp. 2
	Chaetosphaeridium globosum	Cosmarium speciosum	
	Stigeoclonium tenue	Cosmarium of. tenue	Division: Bacillariophyceae
	Ulothrix zonata	Cosmarium tinctum	(Diatoms)~
Order	: Chlorococcales	Cosmarium tinctum	Order: Centrales
	Ankistrodesmus falcatus	v. subretusum	Suborder: Discineae
	v. mirabilis	Cosmarium trilobatum	Family: Coscinodiscaceae
	Ankistrodesmus sprialis	v. depressum	Coscinodiscus rothii
	Characium sp.	Cosmarium cf. undulatum	v. subsalsa
	Chlorella vulgaris	v. minutum	. Cyclotella cf. glomerata
	Chlorococcum humicola	Cosmarium variolatum	Cyclotella meneghiniana
	Coelastrum sp.	Cosmarium species #1-19	Melosira granulata
	Crucigenia tetrapedia	Cylindrocystis spp.	Melosira varians
	Dimorphococcus lunatus	Desmidium aptogonum	Thalassiosira fluviatilis
	Kentrosphaera gloephila	Desmidium baileyi	Order: Pennales
	Kirchneriella lunaris	Desmidium coarctatum	Suborder: Araphidae
	Nephrocytium lunatum	Desmidium cylindricum	Fragilaria brevistriata
			itagilal la Dicristitata
		Desmidium orevelli	
	Oocystis spp. Pediastrum borvanum	Desmidium grevelli Desmidium swartzii	Fragilaria capucina v. mesolepta
	Pediastrum boryanum	Desmidium swartzii	Fragilaria capucina v. mesolepta
	Pediastrum boryanum Pediastrum sp. 1	Desmidium swartzii Docidium undulatum	Fragilaria capucina v. mesolepta Fragilaria pinnata
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans	Desmidium swartzii Docidium undulatum v. semiundulatum	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus	Desmidium šwartzii Docidium undulatum v. semiundulatum Evastrum abruptum	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov.
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum affine	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis	Desmidium swartzii Docidium undulatum v. semiundulatum Evastrum abruptum f. minor Euastrum affine Euastrum bidentatum	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula	Desmidium swartzii Docidium undulatum v. semiundulatum Evastrum abruptum f. minor Evastrum affine Euastrum bidentatum Euastrum binale	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum affine Euastrum bidentatum Euastrum binale v. gutwinskii	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus Scenedesmus longus	Desmidium swartzii Docidium undulatum v. semiundulatum Evastrum abruptum f. minor Euastrum affine Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum affine Euastrum bidentatum Euastrum binale v. gutwinskii	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum affine Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus quadricauda	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum affine Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum lutkemulleri	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus quadricauda Selenastrum spp.	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum affine Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum lutkemulleri Euastrum cf. verrucosum	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov.
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus quadricauda Selenastrum spp. Sorastrum sp.	Desmidium swartzii Docidium undulatum v. semiundulatum Evastrum abruptum f. minor Euastrum effine Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum lutkemulleri Euastrum cf. verrucosum v. reductum	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus quadricauda Selenastrum spp. Sorastrum sp. Sphaerocystis sp. 1	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum bidentatum Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum intermedium Euastrum cf. verrucosum v. reductum Genicularia elegans	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov.
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus quadricauda Selenastrum spp. Sorastrum sp. Sphaerocystis sp. 1 Tetraedron caudatum	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum effine Euastrum bidentatum Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum lutkemulleri Euastrum cf. verrucosum v. reductum Genicularia elegans Genicularia sp. 1	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus quadricauda Selenastrum spp. Sorastrum sp. Sphaerocystis sp. 1 Tetraedron caudatum Tetraedron regulare	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum effine Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum intermedium Euastrum intermedium Euastrum cf. verrucosum v. reductum Genicularia elegans Genicularia sp. l Gonatozygon aculeatum	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva Achnanthes hauckiana
· Ondar	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus brasiliensis Scenedesmus incrassatus Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus paraisiensis Scenedesmus langus	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum effine Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum lutkemulleri Euastrum cf. verrucosum v. reductum Genicularia elegans Genicularia sp. 1 Gonatozygon brebissonii	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva Achnanthes inflata
O rder	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus incrassatus Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus quadricauda Selenastrum spp. Sorastrum sp. Sphaerocystis sp. 1 Tetraedron caudatum Tetraedron regulare Tetraedron trigonum : Oedogonales	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum bidentatum Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum lutkemulleri Euastrum intermedium Euastrum cf. verrucosum v. reductum Genicularia elegans Genicularia sp. l Gonatozygon aculeatum Gonatozygon brebissonii Hyalotheca dissiliens	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra ulna v. oxyrhynchus Synedra ulna v. oxyrhynchus Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva Achnanthes inflata Achnanthes minutissima
´ Order	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus prisiensis Scenedesmus parisiensis Scenedesmus prisiensis Scenedesmus rationala Selenastrum spp. Sphaerocystis sp. 1 Tetraedron ragulare Tetraedron trigonum Cedogonales Bulbochaetae sp.	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum bidentatum Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum lutkemulleri Euastrum cf. verrucosum v. reductum Genicularia elegans Genicularia sp. 1 Gonatozygon aculeatum Gonatozygon brebissonii Hyalotheca dissiliens Hyalotheca undulata	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva Achnanthes inflata Achnanthes minutissima Cocconeis placentula
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus p. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus puadricauda Selenastrum spp. Sorastrum spp. Sphaerocystis sp. 1 Tetraedron caudatum Tetraedron regulare Tetraedron trigonum Dedogonales Bulbochaetae sp. Oedogonium species 1, 2 & 3	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum bidentatum Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum lutkemulleri Euastrum intermedium Cuastrum cf. verrucosum v. reductum Genicularia elegans Genicularia sp. l Gonatozygon aculeatum Gonatozygon brebissonii Hyalotheca dissiliens Hyalotheca undulata Mesotaenium sp. l	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva Achnanthes inflata Achnanthes minutissima Cocconeis placentula v. euglypta
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus p. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus quadricauda Selenastrum spp. Sorastrum sp. Sphaerocystis sp. 1 Tetraedron caudatum Tetraedron regulare Tetraedron trigonum Dedogonales Bulbochaetae sp. Oedogonium species 1, 2 & 3 Zygnematales	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum effine Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum intermedium Euastrum intermedium Euastrum intermedium Cuastrum intermedium Cuastrum intermedium Cuastrum cf. verrucosum v. reductum Genicularia elegans Genicularia sp. 1 Gonatozygon aculeatum Gonatozygon brebissonii Hyalotheca undulata Mesotaenium sp. 1 Micrasterias foliaceae	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva Achnanthes inflata Achnanthes minutissima Cocconeis placentula v. euglypta Suborder: Biraphidae
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus brasiliensis Scenedesmus incrassatus Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus obliquus Scenedesmus quadricauda Selenastrum spp. Sorastrum sp. Sphaerocystis sp. 1 Tetraedron caudatum Tetraedron regulare Tetraedron trigonum cedogonales Bulbochaetae sp. Oedogonium species 1, 2 & 3 Zygnematales Mougeotia species 1 & 2	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum bidentatum Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum lutkemulleri Euastrum cf. verrucosum v. reductum Genicularia elegans Genicularia sp. l Gonatozygon aculeatum Gonatozygon brebissonii Hyalotheca dissiliens Hyalotheca undulata Mesotaenium sp. l Micrasterias follaceae Micrasterias pinnatifida	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva Achnanthes inflata Achnanthes minutissima Cocconeis placentula v. euglypta
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus p. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus prisiensis Scenedesmus incrassatus Scenedesmus prisiensis Scenedesmus incrassatus Scene	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum bidentatum Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum lutkemulleri Euastrum cf. verrucosum v. reductum Genicularia elegans Genicularia sp. 1 Gonatozygon aculeatum Gonatozygon brebissonii Hyalotheca dissiliens Hyalotheca undulata Mesotaenium sp. 1 Micrasterias foliaceae Micrasterias pinnatifida Micrasterias radiata	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva Achnanthes inflata Achnanthes minutissima Cocconeis placentula v. euglypta Suborder: Biraphidae Amphipleura pellucida
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus brasiliensis Scenedesmus brasiliensis Scenedesmus incrassatus Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus quadricauda Selenastrum spp. Sorastrum spp. Sorastrum sp. Sphaerocystis sp. 1 Tetraedron caudatum Tetraedron trigonum Codogonales Bulbochaetae sp. Oedogonium species 1, 2 & 3 Zygnematales Mougeotia species 1 & 2 Spirogyra spp. Families: Mesotaeniaceae	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum fine Euastrum bidentatum Euastrum bidentatum Euastrum bidentatum Euastrum bidente v. gutwinskii Euastrum bidele v. elobatum Euastrum intermedium Euastrum lutkemulleri Euastrum cf. verrucosum v. reductum Genicularia elegans Genicularia sp. l Gonatozygon aculeatum Gonatozygon brebissonii Hyalotheca dissiliens Hyalotheca undulata Mesotaenium sp. l Micrasterias foliaceae Micrasterias pinnatifida Micrasterias radiata Micrasterias radiosa	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva Achnanthes inflata Achnanthes minutissima Cocconeis placentula v. euglypta Suborder: Biraphidae Amphipleura pellucida Amphora veneta
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus p. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus brasiliensis Scenedesmus brasiliensis Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus quadricauda Selenastrum spp. Sorastrum spp. Sorastrum sp. Sphaerocystis sp. 1 Tetraedron caudatum Tetraedron regulare Tetraedron trigonum Oedogonales Bulbochaetae sp. Oedogonium species 1, 2 & 3 Zygnematales Mougeotia species 1 & 2 Spirogyra spp. Families: Mesotaeniaceae and Desmidiaceae	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum affine Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum intermedium Euastrum intermedium Euastrum intermedium Euastrum fo. verrucosum v. reductum Genicularia elegans Genicularia sp. l Gonatozygon aculeatum Gonatozygon brebissonii Hyalotheca undulata Mesotaenium sp. l Micrasterias foliaceae Micrasterias foliaceae Micrasterias radiata Micrasterias radiosa v. ornata	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva Achnanthes inflata Achnanthes minutissima Cocconeis placentula v. euglypta Suborder: Biraphidae Amphora yeneta Amphora sp. 1 Anomoeoneis serians
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum bidentatum Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum lutkemulleri Euastrum cf. verrucosum v. reductum Genicularia elegans Genicularia sp. l Gonatozygon aculeatum Gonatozygon brebissonii Hyalotheca dissiliens Hyalotheca undulata Mesotaenium sp. l Micrasterias foliaceae Micrasterias radiata Micrasterias radiata V. ornata Micrasterias species 1 & 1	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva Achnanthes inflata Achnanthes minutissima Cocconeis placentula v. euglypta Suborder: Biraphidae Amphora yeneta Amphora sp. 1 Anomoeoneis serians
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus pp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus longus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus prisiensis Scenedesmus p	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum affine Euastrum bidentatum Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum intermedium Euastrum cf. verrucosum v. reductum Genicularia elegans Genicularia sp. 1 Gonatozygon aculeatum Gonatozygon brebissonii Hyalotheca dissiliens Hyalotheca dissiliens Hyalotheca undulata Mesotaenium sp. 1 Micrasterias foliaceae Micrasterias radiata Micrasterias radiata Micrasterias radiosa v. ornata Micrasterias species 1 & Retrium digitus	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva Achnanthes inflata Achnanthes minutissima Cocconeis placentula v. euglypta Suborder: Biraphidae Amphipleura pellucida Amphora veneta Amphora veneta Amphora serians Anomoeoneis serians
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus pp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus quadricauda Selenastrum spp. Sonastrum spp. Sonastrum spp. Sonastrum spp. Tetraedron caudatum Tetraedron regulare Tetraedron trigonum Oedogonales Bulbochaetae sp. Oedogonium species 1, 2 & 3 Zygnematales Mougeotia species 1 & 2 Spirogyra spp. Families: Mesotaeniaceae and Desmidiaceae Arthrodesmus incus v. semilunatus Arthrodesmus validus	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum affine Euastrum bidentatum Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum lutkemulleri Euastrum cf. verrucosum v. reductum Genicularia elegans Genicularia sp. l Gonatozygon aculeatum Gonatozygon brebissonii Hyalotheca dissiliens Hyalotheca undulata Mesotaenium sp. l Micrasterias foliaceae Micrasterias foliaceae Micrasterias radiata Micrasterias radiosa v. ornata Micrasterias species 1 & Setrium digitus Onychonema laeve	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva Achnanthes inflata Achnanthes inflata Achnanthes minutissima Cocconeis placentula v. euglypta Suborder: Biraphidae Amphipleura pellucida Amphora sp. 1 Anomoeoneis serians Anomoeoneis serians v. brachysira
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus pp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus puadricauda Selenastrum spp. Sonastrum spp. Sonastrum sp. Sphaerocystis sp. 1 Tetraedron caudatum Tetraedron regulare Tetraedron trigonum Dedogonales Bulbochaetae sp. Dedogonium species 1, 2 & 3 Zygnematales Mougeotia species 1 & 2 Spirogyra spp. Families: Mesotaeniaceae and Desmidiaceae Arthrodesmus incus v. semilunatus Arthrodesmus validus Bambusina brebissonii	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum affine Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum lutkemulleri Euastrum intermedium Euastrum intermedium Genicularia elegans Genicularia sp. l Gonatozygon aculeatum Gonatozygon brebissonii Hyalotheca dissiliens Hyalotheca undulata Mesotaenium sp. l Micrasterias foliaceae Micrasterias radiata Micrasterias radiata Micrasterias radiata Micrasterias radiosa v. ornata Micrasterias species 1 & Metrium digitus Onychonema laeve v. latum	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva Achnanthes inflata Achnanthes minutissima Cocconeis placentula v. euglypta Suborder: Biraphidae Amphipleura pellucida Amphora veneta Amphora veneta Amphora sp. 1 Anomoeoneis serians v. brachysira Anomoeoneis sphaerophora Anomoeoneis yttrea
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus pp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus quadricauda Selenastrum spp. Sorastrum spp. Sorastrum sp. Sphaerocystis sp. 1 Tetraedron caudatum Tetraedron regulare Tetraedron trigonum Dedogonales Bulbochaetae sp. Oedogonium species 1, 2 & 3 Zygnematales Mougeotia species 1 & 2 Spirogyra spp. Families: Mesotaeniaceae and Desmidiaceae Arthrodesmus incus v. semilunatus Arthrodesmus validus Bambusina brebissonii Closterium acutum	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum bidentatum Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum lutkemulleri Euastrum lutkemulleri Euastrum cf. verrucosum v. reductum Genicularia elegans Genicularia sp. l Gonatozygon aculeatum Gonatozygon brebissonii Hyalotheca dissiliens Hyalotheca dissiliens Hyalotheca undulata Mesotaenium sp. l Micrasterias foliaceae Micrasterias foliaceae Micrasterias radiata Micrasterias radiosa v. ornata Micrasterias species 1 & : Netrium digitus Onychonema laeve v. latum Pleurotaenium minutum	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva Achnanthes inflata Achnanthes minutissima Cocconeis placentula v. euglypta Suborder: Biraphidae Amphipleura pellucida Amphora veneta Amphora sp. 1 Anomoeoneis serians v. brachysira Anomoeoneis sphaerophora
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus pp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus prisiensis Scenedes	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum bidentatum Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum lutkemulleri Euastrum cf. verrucosum v. reductum Genicularia elegans Genicularia sp. 1 Gonatozygon aculeatum Gonatozygon brebissonii Hyalotheca dissiliens Hyalotheca dissiliens Hyalotheca undulata Mesotaenium sp. 1 Micrasterias foliaceae Micrasterias foliaceae Micrasterias radiata Micrasterias radiosa v. ornata Micrasterias species 1 & Retrium digitus Onychonema laeve v. latum Pleurotaenium minutum r	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva Achnanthes inflata Achnanthes inflata Achnanthes minutissima Cocconeis placentula v. euglypta Suborder: Biraphidae Amphipleura pellucida Amphora veneta Amphora veneta Amphora serians v. brachysira Anomoeoneis serians v. brachysira Anomoeoneis sphaerophora Anomoeoneis vitrea Bacillaria parpdoxa
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus pp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus longus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus quadricauda Selenastrum spp. Sorastrum spp. Sorastrum spp. Sorastrum spp. Sorastrum regulare Tetraedron caudatum Tetraedron trigonum Codogonales Bulbochaetae sp. Oedogonium species 1, 2 & 3 Zygnematales Mougeotia species 1 & 2 Spirogyra spp. Families: Mesotaeniaceae and Desmidiaceae Arthrodesmus validus Bambusina brebissonii Closterium acutum Closterium archerianum f. majo	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum affine Euastrum bidentatum Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum lutkemulleri Euastrum cf. verrucosum v. reductum Genicularia elegans Genicularia sp. l Gonatozygon aculeatum Gonatozygon brebissonii Hyalotheca dissiliens Hyalotheca dissiliens Hyalotheca undulata Mesotaenium sp. l Micrasterias foliaceae Micrasterias foliaceae Micrasterias radiata Micrasterias radiata Micrasterias radiosa v. ornata Micrasterias species 1 & Metrium digitus Onychonema laeve v. latum Pleurotaenium minutum r elongatum	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva Achnanthes inflata Achnanthes inflata Achnanthes minutissima Cocconeis placentula v. euglypta Suborder: Biraphidae Amphora sp. 1 Anomoeoneis serians Anomoeoneis serians v. brachysira Anomoeoneis sphaerophora Anomoeoneis vitrea Baciilaria parodoxa Caloneis bacillum
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus pp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus incrassatus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus parisiensis Scenedesmus prisiensis Scenedes	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum bidentatum Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum lutkemulleri Euastrum lutkemulleri Euastrum cf. verrucosum v. reductum Genicularia elegans Genicularia sp. l Gonatozygon aculeatum Gonatozygon brebissonii Hyalotheca dissiliens Hyalotheca dissiliens Hyalotheca undulata Mesotaenium sp. l Micrasterias foliaceae Micrasterias foliaceae Micrasterias radiata Micrasterias radiosa v. ornata Micrasterias species 1 & : Netrium digitus Onychonema laeve v. latum Pleurotaenium minutum r Pleurotaenium minutum r Pleurotaenium minutum	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva Achnanthes minutissima Cocconeis placentula v. euglypta Suborder: Biraphidae Amphipleura pellucida Amphora veneta Amphora sp. 1 Anomoeoneis serians v. brachysira Anomoeoneis sphaerophora Anomoeoneis strea Bacillaria parodoxa Caloneis bacillum Caloneis ventricosa
	Pediastrum boryanum Pediastrum sp. 1 Scenedesmus pp. 1 Scenedesmus abundans Scenedesmus acruatus v. platydisca Scenedesmus bijuga Scenedesmus brasiliensis Scenedesmus denticula Scenedesmus longus Scenedesmus longus Scenedesmus obliquus Scenedesmus parisiensis Scenedesmus quadricauda Selenastrum spp. Sorastrum spp. Sorastrum spp. Sorastrum spp. Sorastrum regulare Tetraedron caudatum Tetraedron trigonum Codogonales Bulbochaetae sp. Oedogonium species 1, 2 & 3 Zygnematales Mougeotia species 1 & 2 Spirogyra spp. Families: Mesotaeniaceae and Desmidiaceae Arthrodesmus validus Bambusina brebissonii Closterium acutum Closterium archerianum f. majo	Desmidium swartzii Docidium undulatum v. semiundulatum Euastrum abruptum f. minor Euastrum affine Euastrum bidentatum Euastrum bidentatum Euastrum binale v. gutwinskii Euastrum binale v. elobatum Euastrum intermedium Euastrum lutkemulleri Euastrum cf. verrucosum v. reductum Genicularia elegans Genicularia sp. l Gonatozygon aculeatum Gonatozygon brebissonii Hyalotheca dissiliens Hyalotheca dissiliens Hyalotheca undulata Mesotaenium sp. l Micrasterias foliaceae Micrasterias foliaceae Micrasterias radiata Micrasterias radiata Micrasterias radiosa v. ornata Micrasterias species 1 & Metrium digitus Onychonema laeve v. latum Pleurotaenium minutum r elongatum	Fragilaria capucina v. mesolepta Fragilaria pinnata Synedra delicatissima Synedra pahokeensis sp. nov. Synedra radians Synedra tenera Synedra ulna v. amphirhynchus Synedra ulna v. obtusa Synedra ulna v. oxyrhynchus Synedra ulna v. oxyrhynchus Synedra sp. 2 sp. nov. Suborder: Monoraphidae Achnanthes exigua v. heterovalva Achnanthes minutissima Cocconeis placentula v. euglypta Suborder: Biraphidae Amphipleura pellucida Amphora veneta Amphora sp. 1 Anomoeoneis serians v. brachysira Anomoeoneis sphaerophora Anomoeoneis strea Bacillaria parodoxa Caloneis bacillum Caloneis ventricosa

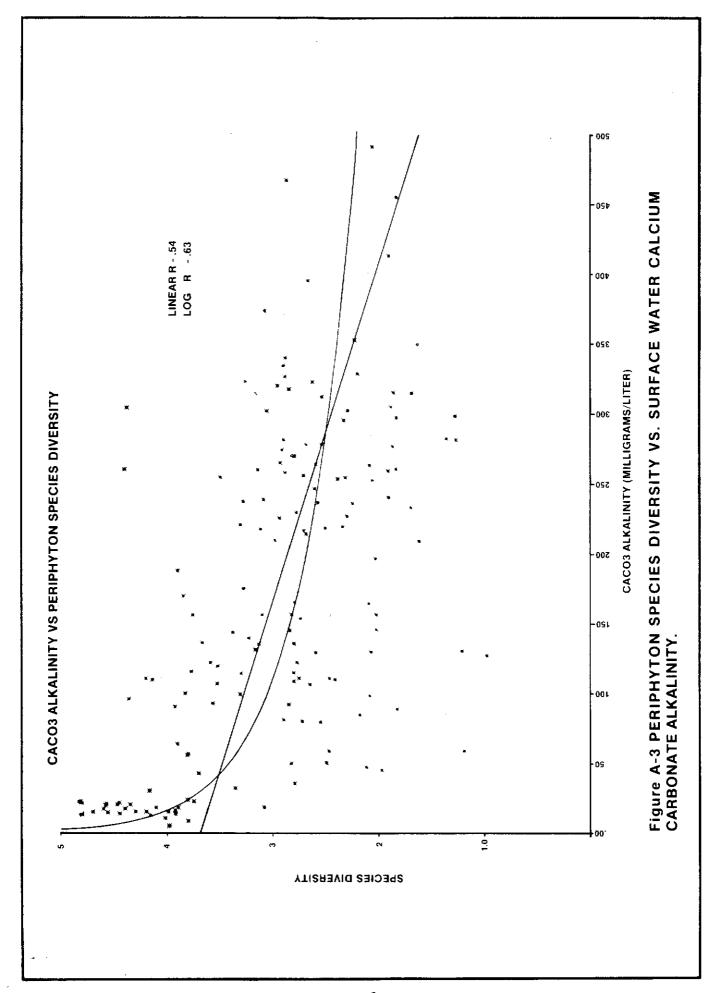
SYSTEMATIC LIST OF PERIPHYTON SPECIES

```
Division: Bacillariophyceae<sup>C</sup> (Con't).
Order: Pennales
                                                     Suborder: Biraphidae (Con't)
                                                          Navicula seminulum
                                                          Navicula cf. subtillissima
      Suborder: Biraphidae
           Cymbella amphioxys
                                                          Navicula tripunctata
                                                          v. schizonemoides
           Cymbella microcephala
            Cymbella minuta
                                                          Navicula variostriata
                                                          Neidium affine
            Cymbella minuta
            v. pseudogracilis
                                                           v. amphirhyncus
            Cymbella minuta
                                                          Neidium cf. iridis
                                                          Nitzschia acicularis
            v. silesiaca
                                                          Nitzschia acuta
            Cymbella of muelleri
           Cymbella pusilla
Cymbella ruttneri
                                                          Nitzschia amphibia
                                                          Nitzschia cf. brevirostris
           Diploneis elliptica
Diploneis finnica
                                                          Nitzschia closterium
                                                          Nitzschia denticula
            Diploneis ovalis
                                                          Nitzschia denticula
            Epithemia spp.
                                                           v. elongata
            Epithemia argus
                                                          Nitzschia dissipata
                                                          Nitzschia fonticola
             v. alpestris
                                                          Nitzschia frustulum
            Eunotia arcus
             v. bindens
                                                          Nitzschia frustulum
                                                           v. perpussilla
            Eunotia curvata
            Eunotia flexuosa
                                                          Nitzschia hybrida
            Eunotia flexuosa
                                                          Nitzschia kutzingiana
                                                          Nitzschia linearis
             v. eurycephala
            Eunotia formica
                                                          Nitzschia microcephala
                                                          Nitzschia obtusa
            Eunotia naegelii
            Eunotia pectinalis
                                                          Nitzschia obtusa
             v. minor
                                                           v. scalpelliformis
            Frustulia rhomboides
                                                          Nitzschia palea
                                                          Nitzschia sigmoidea
            v. crassinervia
            Frustulia rhomboides
                                                          Nitzschia cf. spiculoides
                                                          Nitzschia storchii sp. nov.
             v. saxonica
                                                          Nitzschia cf. subrostrata
            Gomphonema acuminatum
            Gomphonema affine
                                                          Nitzschia subtilis
            Comphonema affine
                                                          Nitzschia tarda
                                                          Nitzschia vermicularis
             v. insigne
                                                          Nitzschia species #5-16
            Gomphonema angustatum
                                                          Pinnularia sp. 2
            Gomphonema consector
            Gomphonema gracile
Gomphonema intricatum
                                                          Pinnularia abaujensis
                                                          Pinnularia acrosphaeria
            Gomphonema intricatum
                                                          Pinnularia biceps
                                                          Pinnularia braunii
             v. vibrio
            Gomphonema subclavatum
                                                           v. amphicephala
            Gomphonema parvulum
                                                          Rhopalodia gibba
                                                          Stauroneis anceps
            Gomphonema truncatum
            Gomphonema turris
                                                           f. gracilis
                                                          Stauroneis phoenicenteron
            Hantzschia amphioxys
                                                           f. gracilis
             v. major
            Mastogloia smithii
                                                          Stauroneis sp. 1
                                                          Stenopterobia intermedia
            Mastogloia smithii
             v. lacustris
                                             Division: Chrysophyceae<sup>D</sup>
            Navicula accomoda
            Navicula arvensis
                                                   Order: Rhizochrysidales
            Navicula bicephala
                                                          Lagynion macrotrachelum
            Navicula biconica
                                                   Order: Rhizochloridates
            Navicula bioculata
                                                          Stipitococcus vasiformes
                                                   Order: Phaeoplacales
            Navicula capitata
                                                          Dinobryon spp.
            Navicula capitata
             v. hungarica
                                                          Epipyxis sp.
                                                                                            Footnote:
            Navicula cincta
                                                          Mallomonas caudata
            Navicula confervacea
                                              Division: Dinophyceae
                                                                                            <sup>a</sup>After Drouet's classification
                                                   Order: Gymnodiniales
            Navicula cryptocephala
                                                                                                 Drouet and Daily (1956);
Drouet, 1968, 1973
             v. veneta
                                                          Gymnodinium sp.
            Navicula cuspidata
Navicula disputans
                                                   Order: Peridinales
                                                          Gonyaulux sp. 1
                                                                                            DAfter G.M. Smith (1950)
            Navicula exigua
                                                          Peridinium species 1 & 2
             y, capitata
                                                                                            CAfter Hustedt's (1930)
            Navicula geitleri
Navicula luzonensis
                                               Division: Euglenophyceae
Order: Euglenales
                                                           Euglena spp.
            Navicula minima
                                                          Phacus spp.
            Navicula mutica
                                                           Trachelomonas spp.
            Navicula paucivisitata
            Navicula pupula
             v. rectangularis
                                                Division: Cryptophyceae
                                                     Order: Cryptomonales
            Navicula pygmaea
                                                           Cryptomonas erosa
            Navicula radiosa
                                                           Rhodomonas spp.
            Navicula radiosa
             v. tenella
            Naviucla rhynchocephla
                                               Division: Charophyceae
                                                     Order: Charales
             v. germainii
```

Chara spp.







APPENDIX B

TAXONOMIC NOTES

Schizothrix calcicola (Ag.) Gom. was the most widely distributed and most frequently encountered blue-green algae observed in our collections, occurring in every sample examined. S. calcicola is a small threadlike filamentous alga consisting of a trichome (usually) enclosed within a thin mucilaginous sheath. Trichome length was highly variable with trichome width averaging 1.2-1.5 (up to 3.5) microns. Average cell volume for S. calcicola ranged between 300-500 cubic microns.

Presently there exists confusion regarding the taxonomy of this alga. Drouet (1968) has reported that strains of Schizothrix can modify their cell morphology (ie. cell size or shape, degree of filament bending, presence or absence or a sheath, or color), in response to changing environmental or physiological conditions. Algae which have these characteristics have been termed "ecophenes". Drouet (1963) demonstrated these morphological variations by growing pure cultures of S. calcicola, in a wide variety of culture media under different environmental conditions. His results showed the final growth form of this one species could be classified as 54 separate species representing nine different genera using some of the most commonly used taxonomic references. Gleason (1972) reports similar "variants" of S. calcicola both in his laboratory culture work and from periphyton mats collected in Everglades National Park. Within this study five ecophenes of S. calcicola are reported.

Scytonema hofmannii was the second most abundant filamentous blue-green present occurring at times as a co-dominant species in association with Schizothrix calcicola at calcium rich interior marsh sites. Filaments of S. hofmannii were much larger in size (roughly 500 times) as compared to S. calcicola with a cell diameter typically ranging between 8-12 (up to 25) microns. Cell volumes calculated for S. hofmannii averaged between 150,000-250,000 cubic microns. Scytonema hofmannii is identified by its false branching habit, yellow-brown colored trichomes, intercalary heterocysts, and laminated hyaline sheath that are usually heavily encrusted with calcium carbonate crystals.

S. hofmannii was a relatively slow growing species and was not as commonly encountered in our glass slide collections as compared to periphyton mat cores collected from marsh sediments. S. hofmannii on glass slide substrates accounted for a maximum of 45 percent of the total periphyton community (by volume)

whereas <u>S. hofmannii</u> accounted for 50-80 percent of the algae collected from periphyton mat cores. <u>S. hofmannii</u> and <u>S. calcicola</u> were the dominant periphyton flora in waters containing high concentrations of dissolved minerals (bicarbonate and calcium) and low concentrations of inorganic nutrients.

Microcoleus lyngbyaceus was the most common blue-green present at nutrient enriched peripheral marsh sites. M. lyngbyaceus is a relatively large filamentous blue-green intermediate in size between S. calcicola and S. hofmanni. It is distinguished by its parallel trichomes, and highly granular cross walls. Sheath material may be present or completely lacking. Living specimens are mobile and exhibit a gliding motion under the microscope.

The taxonomy of M lyngbyaceus is also in a muddled state. Drouet's (1968) revision of this group has reduced or "lumped" hundreds of filamentous blue-green species into six genera and 23 species. As a result, taxonomists using earlier reference material (eg. Prescott, 1962; G.M. Smith, 1950) could easily key out M. lyngbyaceus as one of five different species depending upon cell diameter measurements or the presence or absence of sheath material. For example; Drouet (1968) recognizes Oscillatoria limosa, Oscillatoria tenius, Lyngbya major, Lyngbya martensiana and Lyngbya aestuarii as being merely ecophenes of Microcoleus lyngbyaceus. It is not the intention of this paper to address these problems of classical taxonomy, but rather to report that M. lyngbyaceus commonly exhibited a wide range of morphological variability within our collections. It should be noted, however, that numerous studies have cited Oscillatoria limosa and Oscillatoria tenuis (= M. lynbyaceus) as two of the most pollution tolerant algae reported in the literature. Large populations of these algae have been cited as indicators of gross organic pollution (Palmer, 1969).

Five genera of filamentous greens were quantitatively important members of the WCA periphyton community. These included: Mougeotia, Spirogyra, Bulbochaetae, Oedogonium, and Stigeoclonium. Mougeotia was the most common and widely distributed of these genera. Both Mougeotia and Spirogyra are members of the family Zygnematales. These unbranched, filamentous greens reproduce sexually by use of a conjugation tube connected between two filaments whereby gametes are exchanged to form zygospores (Smith, 1950;

Whitford and Schmuacher, 1969). Members of this family are difficult to identify to species as most manuals require the presence of fruiting bodies (ie. zygospores). The majority of filamentous greens colonizing our slides were in the vegetative stage and were only keyed to genus.

Oedogonium and Bulbochaetae were also important components of the WCA periphyton community. Oedogonium spp. is an unbranched,

filamentous green algae which is identified by the presence of one or more ring-like scars (resulting from previous cell division) located at the enlarged anterior end of the cell. Oedogonium is difficult to key to species as both male and female reproductive structures are required for identification. Since the majority of filaments colonizing our slides were in the vegetative stage, no attempt was made to key these organisms lower than the generic level.